



City Research Online

City St George's, University of London

Citation: Roberts, P. A. (2022). Mathematical models of retinitis pigmentosa: The trophic factor hypothesis. *Journal of Theoretical Biology*, 534, 110938. doi: 10.1016/j.jtbi.2021.110938

This is the accepted version of the paper.

This version of the publication may differ from the final published version. To cite this item please consult the publisher's version.

Permanent repository link: <https://openaccess.city.ac.uk/id/eprint/33409/>

Link to published version: <https://doi.org/10.1016/j.jtbi.2021.110938>

Copyright and Reuse: Copyright and Moral Rights remain with the author(s) and/or copyright holders. Copies of full items can be used for personal research or study, educational, or not-for-profit purposes without prior permission or charge, unless otherwise indicated, provided that the authors, title and full bibliographic details are credited, a hyperlink and/or URL is given for the original metadata page and the content is not changed in any way. For full details of reuse please refer to [City Research Online policy](#).

Mathematical Models of Retinitis Pigmentosa: The Trophic Factor Hypothesis

Paul A. Roberts^{a,*}

^a*School of Life Sciences, University of Sussex, John Maynard Smith Building, Brighton, BN1 9QG, UK*

Abstract

Retinitis pigmentosa (RP) is the term used to denote a group of inherited retinal-degenerative conditions that cause progressive sight loss. Individuals with this condition lose their light-sensitive photoreceptor cells, known as rods and cones, over a period of years to decades; degeneration starting in the retinal periphery, and spreading peripherally and centrally over time. RP is a rod-cone dystrophy, meaning that rod health and function are affected earlier and more severely than that of cones. Rods degenerate due to an underlying mutation, whereas the reasons for cone degeneration are unknown. A number of mechanisms have been proposed to explain secondary cone loss and the spatio-temporal patterns of retinal degeneration in RP. One of the most promising is the trophic factor hypothesis, which suggests that rods produce a factor necessary for cone survival, such that, when rods degenerate, cone degeneration follows. In this paper we formulate and analyse mathematical models of human RP under the trophic factor hypothesis. These models are constructed as systems of reaction-diffusion partial differential equations in one spatial dimension, and are solved and analysed using a combination of numerical and analytical methods. We predict the conditions under which cones will degenerate following the loss of a patch of rods from the retina, the critical trophic factor treatment rate required to prevent cone degeneration following rod loss and the spatio-temporal patterns of cone loss that would result if the trophic factor mechanism alone were responsible for retinal degeneration.

Keywords: Partial Differential Equations, Asymptotic Analysis, Retina, Photoreceptors, Rod-derived Cone Viability Factor

Declarations of interest: none.

1. Introduction

The group of inherited retinal diseases, known collectively as retinitis pigmentosa (RP), have been the subject of many decades of research. Despite this attention, many of the mechanisms underpinning RP have yet to be conclusively determined, though a number have been hypothesised (Hamel, 2006; Hartong et al., 2006). In this paper, we formulate mathematical models to describe and predict retinal degeneration in human RP for one of the leading hypotheses: the trophic factor hypothesis.

*Corresponding author

Email address: p.a.roberts@univ.oxon.org (Paul A. Roberts)

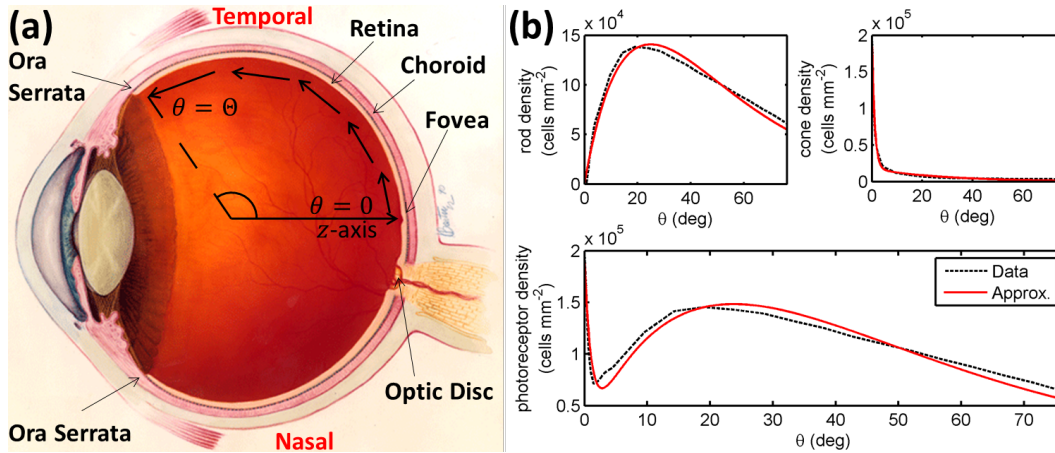


Figure 1: Diagrams of the human eye and retinal photoreceptor distribution (reproduced, with permission, from Roberts et al., 2017). (a) Diagram of the (right) human eye, viewed in the transverse plane, illustrating the model geometry. All models are posed on a domain spanning the region between the foveal centre, at $\theta = 0$, and the ora serrata, at $\theta = \Theta$, along the temporal horizontal meridian, where θ measures the eccentricity. Figure originally reproduced, with modifications, from <http://www.nei.nih.gov/health/coloboma/coloboma.asp>, courtesy: National Eye Institute, National Institutes of Health (NEI/NIH). (b) Measured and fitted photoreceptor profiles, along the temporal horizontal meridian, in the human retina. Cone profile: $\tilde{p}_c(\theta) = B_1 e^{-b_1 \theta} + B_2 e^{-b_2 \theta}$, and rod profile: $\tilde{p}_r(\theta) = B_3 \theta e^{-b_3 \theta}$ (see Table 1 for parameter values). The photoreceptor profile is the sum of the rod and cone profiles ($\tilde{p}_r(\theta) + \tilde{p}_c(\theta)$). Experimental data provided by Curcio and published in Curcio et al. (1990).

The retina is the tissue layer at the back of the eye responsible for light detection (Fig. 1(a)). Retinal light-detecting cells, known as photoreceptors, come in two varieties: rods and cones. Rods confer peripheral, achromatic vision under low-light (scotopic) conditions, while cones confer high-acuity central, chromatic vision under well-lit (photopic) conditions (Oyster, 1999). In the human retina, considered here, cones are mostly concentrated in the centre of the retina, known as the fovea, whereas rods dominate the mid- and far-peripheral retina (Curcio et al., 1990) (Fig. 1(b)). This is in contrast to the retinas of model animal RP species, such as mice (Ortín-Martínez et al., 2014), rats (Gaillard et al., 2009) and pigs (Chandler et al., 1999), which lack a fovea, their rod and cone densities varying more gradually across the eye (see Section 5 for discussion). Both rods and cones are composed of inner and outer segments, together with an axon, ending in a synaptic terminal. Outer segments (OSs) contain a series of discs in which light-sensitive photopigments are embedded (Oyster, 1999). Each day, OS tips are shed and phagocytosed by the underlying retinal pigment epithelium (RPE), while new discs are generated where the OS meets the inner segment (Guérin et al., 1993; Jonnal et al., 2010, 2012; Kocaoglu et al., 2016; Pircher et al., 2011; Young, 1967, 1971, 1978; Young and Bok, 1969).

RP is a rod-cone dystrophy, meaning that rod function and health are affected earlier and more severely than cone function and health (though there do exist rarer cone-rod dystrophy forms, and forms in which rods and cones are affected on a similar time scale, Hamel, 2006; Hartong et al., 2006). This is often because it is the rods, rather than the cones, which express the disease-causing mutation (though mutations can alternatively be expressed in the RPE, Daiger et al., 2007; Hamel, 2006; Hartong et al., 2006). Retinal degeneration (and hence visual field loss) typically initiates in the mid- or far-periphery of the retina, starting with small patches of photoreceptor loss (Cideciyan et al., 1998; García-Ayuso et al., 2013; Ji et al., 2012; Lee et al., 2011; Zhu et al., 2013) which spread and coalesce into larger regions, causing blind spots

(scotomas). Degeneration spreads through the rod-dominated mid- and far-periphery until all peripheral vision is lost (tunnel vision), finally spreading to the cone-rich central retina (macula), leading to the loss of central vision, and hence, total blindness (Grover et al., 1998; Hamel, 2006; Hartong et al., 2006). RP is the most common inherited retinal degeneration, with a prevalence of approximately 1 in 4000, and is presently untreatable (Hamel, 2006; Hartong et al., 2006; Musarella and MacDonald, 2011; Shintani et al., 2009).

Given that the disease-causing mutations are not typically expressed by cones, it is unclear what causes them to degenerate. Further, the reasons behind the spatio-temporal patterns of retinal degeneration in humans (who have a sharper spatial variation in rod and cone densities than in RP animal models), and for which any particular degenerate patch expands or remains stable in humans or animal models, remain a mystery (Grover et al., 1998; Hamel, 2006; Hartong et al., 2006). A number of mechanisms have been proposed to explain secondary cone loss, which also have important implications for spatio-temporal patterns and degenerate patch expansion, most notably trophic factor (TF) depletion (Aït-Ali et al., 2015; Léveillard et al., 2004; Mei et al., 2016), oxygen toxicity (Stone et al., 1999; Travis et al., 1991; Valter et al., 1998), metabolic dysregulation (Punzo et al., 2009, 2012), toxic substances (Ripps, 2002) and microglia (Gupta et al., 2003). In this paper, we consider the first of these, often denoted as the trophic factor hypothesis. This hypothesis suggests that rods produce a TF necessary for cone survival, such that, when rods degenerate, the TF is depleted, and cone degeneration follows. Such a factor, known as rod-derived cone viability factor (RdCVF), has been chemically identified by Léveillard et al. (2004), and shown to slow cone degeneration and preserve cone function in chick, rat and mouse models (Fintz et al., 2003; Léveillard et al., 2004; Mohand-Saïd et al., 1998, 2000, 1997; Yang et al., 2009). RdCVF promotes cone survival by binding to the photoreceptor transmembrane protein Basigin-1 (BSG1), which in turn binds to the glucose transporter GLUT1, increasing cone glucose uptake and stimulating aerobic glycolysis (Aït-Ali et al., 2015). Further, while produced by both rods and cones, the presence of neighbouring rods is required for the production of RdCVFL (RdCVF long) in cones, an enzyme which protects against oxidative stress, while RdCVF acts upstream of RdCVFL to aid in its reduction (Léveillard and Sahel, 2017; Mei et al., 2016; Reichman et al., 2010). As such, it has been speculated that cones may ultimately die in RP due to oxidative damage following loss of (reduced) RdCVFL (Léveillard and Sahel, 2017). For reviews, see Clérin et al. (2020); Léveillard and Aït-Ali (2017); Léveillard and Sahel (2010, 2017); Mohand-Saïd et al. (2001).

In this paper, we construct mathematical models to describe and predict retinal degeneration and preservation in humans under the trophic factor hypothesis. As such, we are isolating the TF (RdCVF) mechanism, in a manner that would be almost impossible to achieve experimentally, to investigate what would happen if this mechanism were solely responsible for cone degeneration in RP. We do not mean to claim by this that RdCVF starvation is *in fact* solely responsible for cone degeneration, rather, we are asking the hypothetical question ‘*what if* RdCVF starvation were solely responsible for cone degeneration?’. Further, while not explicitly modelled, our models are consistent with the ultimate cause of cone death being oxygen toxicity, where the loss of RdCVF leads to the loss of reduced RdCVFL (both due to the loss of RdCVF’s reducing power and, possibly, as Mei et al. speculate, due to the necessity of the presence for RdCVF for RdCVFL production in cones, Mei et al., 2016). Our models are formulated as systems of reaction-diffusion partial differential equations (PDEs) in one spatial dimension, and solved and analysed using a combination of numerical and analytical techniques. We use these models to address three key questions: (i) under what conditions will the loss of a patch of rods lead to cone degeneration?, (ii) what is the critical TF treatment dose required to prevent cone degeneration following the loss of a patch of rods?, and (iii) what spatio-temporal patterns of cone degeneration can be explained via the TF mechanism alone?

In earlier work, we developed the first and only models to consider the oxygen toxicity hypothesis, and to take the key step of accounting for the distribution of photoreceptors (Roberts et al., 2017, 2018, see also, Roberts et al. 2016b). These models are formulated as systems of reaction-diffusion PDEs in one

and two spatial dimensions, and were used to answer the long-standing question as to the spatio-temporal patterns of retinal degeneration that would result from this mechanism alone. These studies further predicted the propagation speed of wavefronts of hyperoxia-driven retinal degeneration, the conditions under which degenerate patches will expand or remain stable, and the effects of treatment with antioxidants and trophic factors.

Burns *et al.* (2002) developed the only (1D PDE) model to consider the toxic substance hypothesis. This model captures the patchy loss of photoreceptors seen in the early stages of RP and, under the right conditions, replicates the exponential decline in photoreceptors observed by Clarke *et al.* (2000, 2001).

Lastly, Camacho *et al.* have produced a series of ordinary differential equation models of the trophic factor hypothesis, which focus on the biochemical details, rather than the spatial spread, of retinal degeneration (Colón Vález *et al.*, 2003; Camacho *et al.*, 2010; Camacho and Wirkus, 2013; Camacho *et al.*, 2014, 2016a,b,c, 2019, 2020, 2021; Wifvat *et al.*, 2021). Their models replicate the rhythmic shedding and renewal of photoreceptor OSs; predict possible disease stages through which a retina may pass towards total blindness, recapitulating known RP phenotypes; identify key reactions and processes; and predict optimum treatment strategies. The work presented here is complementary to Camacho *et al.*'s models, focusing on the spatial spread of degeneration, while remaining relatively agnostic as to the underlying biochemical details (and hence is compatible with a range of TF-related biochemical mechanisms). For further details on the mathematical modelling of the retina, including RP, see our review: Roberts *et al.* (2016a).

The models developed and analysed in this paper are the first trophic factor hypothesis models to be spatially-resolved, or to account for photoreceptor distribution. As such, these are the first TF models capable of answering the three key questions posed above (though we note that Camacho *et al.*'s work considers spatially-uniform analogues of questions (i) and (ii), see especially, Camacho *et al.*, 2014, 2020).

The remainder of this paper is structured as follows. In Section 2, we formulate and non-dimensionalise our mathematical models. In Section 3, we consider the steady-state model, performing asymptotic analyses (Section 3.1), and comparing numerical and analytical solutions (Section 3.2) in the untreated (Section 3.2.1) and treated (Section 3.2.2) cases. In Section 4, we consider numerical solutions to the full dynamic (time dependent) model, in both the untreated (Section 4.1) and treated (Section 4.2) cases. Lastly, in Section 5, we discuss our results and suggest directions for future research.

2. Model formulation

We begin by formulating a model to describe the photoreceptor and TF dynamics within the human retina. Given that the retina lines the essentially spherical inner surface of the eye, we pose our model in spherical polar coordinates (r, θ, ϕ) , with origin at the centre of the vitreous body and orientated such that the z -axis passes outwards through the centre of the fovea (see Fig. 1(a)). We make two geometrically simplifying assumptions. First, we assume that the retina is symmetric about the z -axis, such that the azimuthal dimension, ϕ (rad), can be neglected. While this ignores the optic disc (from which photoreceptors are absent) and other inhomogeneities in rod and cone densities, this is a reasonable simplification to make in the first spatial model of this disease mechanism. Second, since the retina's width ($\sim 80\text{--}320\ \mu\text{m}$, Webvision, <https://webvision.med.utah.edu/>) is approximately two orders of magnitude smaller than its radius of curvature ($\sim 1.2\ \text{cm}$, Oyster, 1999), we depth-average through the retina, assuming a fixed radius $r = R$ (m). Thus, we have reduced the geometry to (R, θ) ; posing our model on the one-dimensional domain $\theta \in [0, \Theta]$ (rad), where $\theta = 0$ (rad) is located at the foveal centre and $\theta = \Theta$ (rad) is located at the ora serrata.

We consider two scenarios, leading to alternate forms of the governing equations:

- Scenario 1: without (w/o) cone regeneration;

- Scenario 2: with cone regeneration.

In Scenario 1, we model the cone photoreceptor density, $p_c(\theta, t)$ (photoreceptors m^{-2}), with its associated initial value, $p_{c_{\text{init}}}(\theta)$ (photoreceptors m^{-2}). In this case, cone degeneration corresponds to the loss of whole cone photoreceptor cells. Thus, given that photoreceptors cannot be regrown once they are lost (i.e. when both inner and outer segments are missing; Zhang et al., 2021), recovery is impossible. In Scenario 2, we model the local cone OS length, $p_c^L(\theta, t)$ (m), with its associated initial value, $p_{c_{\text{init}}}^L(\theta)$ (m). In this case, cone degeneration corresponds to the shortening of cone OS, while leaving the rest of the photoreceptor intact, such that cone density remains fixed at $p_{c_{\text{init}}}(\theta)$ (photoreceptors m^{-2}) for all time. As such, cone regeneration is possible in the form of OS regrowth (see Gu erin et al., 1993, and also Chrysostomou et al., 2008; Lee et al., 2012). Since prolonged exposure to adverse conditions will lead to the eventual loss of the whole photoreceptor, it is assumed in this scenario that adverse conditions are either mild or relatively brief.

We construct a system of PDEs in terms of the dependent variables: TF concentration, $f(\theta, t)$ (M), rod photoreceptor density, $p_r(\theta, t)$ (photoreceptors m^{-2}), and cone photoreceptor density, $p_c(\theta, t)$ (photoreceptors m^{-2}), or local cone OS length, $p_c^L(\theta, t)$ (m); as functions of the independent variables: polar angle, θ (rad), and time, t (s), where $t > 0$ (s).

The TF equation takes the following form,

$$\frac{\partial f}{\partial t} = \underbrace{\frac{D_f}{R^2 \sin(\theta)} \frac{\partial}{\partial \theta} \left(\sin(\theta) \frac{\partial f}{\partial \theta} \right)}_{\text{diffusion}} + \underbrace{\alpha p_r}_{\text{production}} + \underbrace{\xi T(\theta, t)}_{\text{treatment}} - \underbrace{\beta C(\theta, f, p_c)}_{\text{consumption}} - \underbrace{\eta f}_{\text{decay}}, \quad (1)$$

where R (m), the radial position of the retina, D_f ($\text{m}^2 \text{s}^{-1}$), the TF diffusivity, α ($\text{Mm}^2 \text{photoreceptors}^{-1} \text{s}^{-1}$), the rate of TF production by rods, ξ (M s^{-1}), the rate of supply of TF through treatment, β ($\text{m}^2 \text{photoreceptors}^{-1} \text{s}^{-1}$), the rate of TF consumption by cones, and η (s^{-1}), the rate of TF decay, are positive constants. We define the Heaviside step function, $H(\cdot)$, such that

$$H(x) := \begin{cases} 0 & \text{if } x < 0, \\ 1 & \text{if } x \geq 0. \end{cases}$$

The treatment function is defined as follows

$$T(\theta, t) = \begin{cases} H(t - t_{\text{crit}}) & \text{global treatment,} \\ \underbrace{(1 - [H(\theta_{\text{treat}_1} - \theta) + H(\theta - \theta_{\text{treat}_2}])]}_{\text{localisation}} \underbrace{H(t - t_{\text{crit}})}_{\text{initiation}} & \text{local treatment,} \end{cases}$$

where trophic factor (RdCVF) treatment is given from time $t = t_{\text{crit}} > 0$ (s), and can be applied either globally, across the whole domain $\theta \in [0, \Theta]$ (rad), or locally, within the region $\theta \in (\theta_{\text{treat}_1}, \theta_{\text{treat}_2})$ (rad) only. The TF consumption function is defined as follows

$$C(\theta, f, p_c) = \begin{cases} f p_c & \text{w/o cone regeneration,} \\ f p_{c_{\text{init}}}(\theta) & \text{with cone regeneration,} \end{cases}$$

where $p_{c_{\text{init}}}(\theta)$ (photoreceptors m^{-2}), the initial cone density, is defined below. In Scenario 1, cone density may decrease, such that the rate of TF consumption changes both with decreasing $p_c(\theta, t)$ and changing $f(\theta, t)$. By contrast, in Scenario 2, the cone density remains fixed, such that the rate of TF consumption changes with $f(\theta, t)$ only.

The rod equation is as follows

$$\frac{\partial p_r}{\partial t} = \begin{cases} 0 & \text{w/o mutation-induced rod degeneration,} \\ - \underbrace{\phi_r p_r}_{\substack{\text{cell degeneration} \\ \text{(mutation-induced)}}} & \text{with mutation-induced rod degeneration,} \end{cases} \quad (2)$$

where ϕ_r (s^{-1}), the rate of mutation-induced rod degeneration, is a positive constant. We keep the rod density constant in the case where we seek to focus on the photoreceptor dynamics following the formation of a degenerate patch (from which rods and/or cones are absent), while an exponential decline in rods more faithfully represents the natural disease progression. Therefore, in the case without mutation-induced rod degeneration, the rod density remains at its initial value, $p_r(\theta, t) = p_{r_{\text{init}}}(\theta)$ (photoreceptors m^{-2}), while in the case with mutation-induced rod degeneration, $p_r(\theta, t) = p_{r_{\text{init}}}(\theta)e^{-\phi_r t}$ (photoreceptors m^{-2}), provided there is no delay in onset or interruption of degeneration.

In Scenario 1, the cone photoreceptor dynamics are given by

$$\frac{\partial p_c}{\partial t} = \underbrace{\delta p_c \lambda_2(f)}_{\substack{\text{cell degeneration} \\ \text{(TF starvation)}}, \quad (3)$$

where δ (s^{-1}), the rate of TF starvation-induced cone degeneration, is a positive constant.

In Scenario 2, the cone photoreceptor dynamics are given by

$$\frac{\partial p_c^L}{\partial t} = \left[\underbrace{\mu_1 \lambda_3(p_c^L)}_{\substack{\text{OS regrowth} \\ \text{(phase 1)}}} + \underbrace{\mu_2 \left(1 - \frac{p_c^L}{\tilde{p}_c^L}\right) \lambda_4(p_c^L)}_{\substack{\text{OS regrowth} \\ \text{(phase 2)}}} \right] \lambda_1(f) \lambda_5(p_{c_{\text{init}}}) - \underbrace{\delta_L \lambda_2(f) \lambda_6(p_c^L)}_{\substack{\text{OS degeneration} \\ \text{(TF starvation)}}, \quad (4)$$

where \tilde{p}_c^L (m), the healthy cone OS length, μ_1 (s^{-1}), the growth rate of cone OS in phase 1, μ_2 (s^{-1}), the growth rate of cone OS in phase 2, and δ_L (ms^{-1}), the rate of TF starvation-induced cone OS degeneration, are positive constants. In Scenario 1, cone cells degenerate exponentially, while in Scenario 2, individual cone OS degenerate at a constant rate (assuming OS regeneration ceases and an equal length of OS is shed daily, see the Supplementary Materials for further details).

Model fitting to cone OS regrowth data from Guérin et al. (1993) reveals that cone OS regrowth is well-described by a two-phase model. Phase 1, constant growth, occurs for cone OS lengths between 0 and 0.33 as a proportion of their full length. In this early, rapid regrowth phase, new OS discs are regenerated, but OS shedding does not occur. Phase 2, hyperbolic growth, occurs for cone OS lengths between 0.33 and 1 as a proportion of their full length. In this later, slower regrowth phase, new OS discs continue to be regenerated, while OS shedding is re-established. The functions λ_{1-6} are given by

$$\begin{aligned} \lambda_1(f) &= H(f - f_{\text{crit}}), \\ \lambda_2(f) &= 1 - \lambda_1(f) = 1 - H(f - f_{\text{crit}}), \\ \lambda_3(p_c^L) &= H(0.33\tilde{p}_c^L - p_c^L), \\ \lambda_4(p_c^L) &= 1 - \lambda_3(p_c^L) = 1 - H(0.33\tilde{p}_c^L - p_c^L), \\ \lambda_5(p_{c_{\text{init}}}) &= 1 - H(-p_{c_{\text{init}}}), \\ \lambda_6(p_c^L) &= 1 - H(-p_c^L), \end{aligned}$$

where f_{crit} (M), the TF threshold concentration, is a positive constant.

The functions $\lambda_1(f)$ and $\lambda_2(f)$ determine when the cone regrowth and degeneration terms are active. Cone degeneration occurs when $f < f_{\text{crit}}$ (TF starvation) in both Scenarios 1 and 2, while cone (OS) regrowth occurs when $f \geq f_{\text{crit}}$ in Scenario 2, where TF levels are sufficient to maintain cones in health. The functions $\lambda_3(p_c^L)$ and $\lambda_4(p_c^L)$ determine whether phase 1 or phase 2 cone OS regrowth is active. Phase 1 is active when $p_c^L(\theta, t) \leq 0.33\tilde{p}_c^L$, while phase 2 is active when $p_c^L(\theta, t) > 0.33\tilde{p}_c^L$. The function $\lambda_5(p_{c_{\text{init}}})$ prevents cone OS regrowth in regions from which cones are absent initially. The function $\lambda_6(p_c^L)$ halts cone OS degeneration when their length reaches zero, preventing cone OS length becoming negative.

Lastly, we close the system by imposing boundary and initial conditions. We impose zero-flux boundary conditions at both the fovea ($\theta = 0$) and ora serrata ($\theta = \Theta$):

$$\frac{\partial f}{\partial \theta}(0, t) = 0 = \frac{\partial f}{\partial \theta}(\Theta, t). \quad (5)$$

A zero-flux condition is required by symmetry at $\theta = 0$, while the zero-flux condition at $\theta = \Theta$ is justified by the assumption, in the absence of experimental data on TF flux, that TF cannot escape from the retina at the ora serrata.

We impose the following initial conditions:

$$\begin{aligned} f(\theta, 0) &= f_{\text{init}}(\theta), & p_r(\theta, 0) &= p_{r_{\text{init}}}(\theta) = F(\theta)\tilde{p}_r(\theta), \\ p_c(\theta, 0) &= p_{c_{\text{init}}}(\theta) = G(\theta)\tilde{p}_c(\theta), & p_c^L(\theta, 0) &= p_{c_{\text{init}}}^L(\theta) = G(\theta)\tilde{p}_c^L, \end{aligned} \quad (6)$$

where $f_{\text{init}}(\theta)$ (M) is the steady-state solution to Eqs. (1) and (5) with $p_r = p_{r_{\text{init}}}(\theta)$ (photoreceptors m^{-2}), $p_c = p_{c_{\text{init}}}(\theta)$ (photoreceptors m^{-2}) and $T(\theta, 0) = 0$ (no treatment), while $\tilde{p}_r(\theta)$ (photoreceptors m^{-2}), the healthy rod distribution, and $\tilde{p}_c(\theta)$ (photoreceptors m^{-2}), the healthy cone distribution are defined as

$$\begin{aligned} \tilde{p}_r(\theta) &= B_3\theta e^{-b_3\theta}, \\ \tilde{p}_c(\theta) &= B_1e^{-b_1\theta} + B_2e^{-b_2\theta}, \end{aligned}$$

where B_1 (photoreceptors m^{-2}), B_2 (photoreceptors m^{-2}), B_3 (photoreceptors $\text{m}^{-2} \text{rad}^{-1}$), b_1 (rad^{-1}), b_2 (rad^{-1}) and b_3 (rad^{-1}) are positive constants, obtained by fitting to physiological data (see Fig. 1(b), Fig. 2(a) and Curcio et al., 1990). We make the simplifying assumption that healthy cone OS length, \tilde{p}_c^L (m), is constant across the retina. We consider two types of initial conditions for rods and cones: healthy and containing a degenerate patch. Which of these situations obtain is determined by the functions $F(\theta)$ and $G(\theta)$, which are given by

$$\begin{aligned} F(\theta) &= \begin{cases} 1 & \text{if rods healthy initially,} \\ H(\theta_{r_1} - \theta) + H(\theta - \theta_{r_2}) & \text{if rods absent from } \theta \in (\theta_{r_1}, \theta_{r_2}), \end{cases} \\ G(\theta) &= \begin{cases} 1 & \text{if cones healthy initially,} \\ H(\theta_{c_1} - \theta) + H(\theta - \theta_{c_2}) & \text{if cones absent from } \theta \in (\theta_{c_1}, \theta_{c_2}), \end{cases} \end{aligned}$$

where θ_{r_1} (rad) and θ_{r_2} (rad) are the left- (central) and right-hand (peripheral) boundaries of a patch of rod degeneration, and θ_{c_1} (rad) and θ_{c_2} (rad) are the left- (central) and right-hand (peripheral) boundaries of a patch of cone degeneration. Example initial conditions are presented in Fig. 2(b)–(d).

Parameter values associated with the dimensional model can be found in Table 1. See the Supplementary Materials for justification of parameter values.

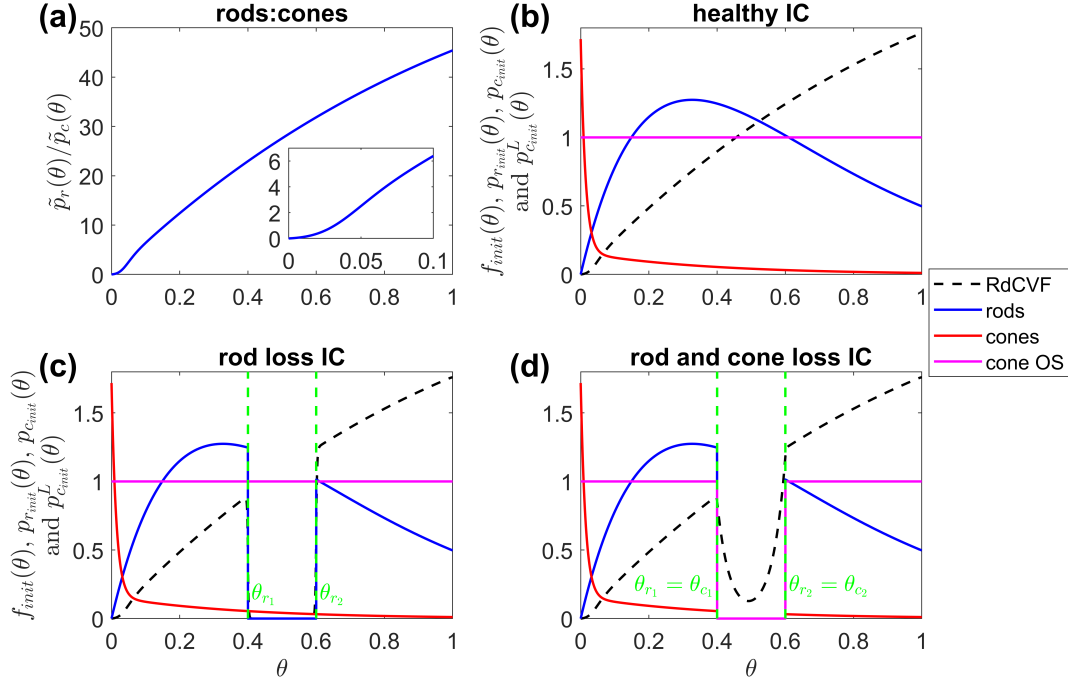


Figure 2: Ratio of rods to cones and exemplar initial conditions. (a) variation in the healthy rod:cone ratio, $\bar{p}_r(\theta)/\bar{p}_c(\theta)$, with eccentricity (inset shows magnified bottom-left corner). (b)–(d) examples of typical initial conditions (ICs) in the healthy retina (b), with a patch of rod loss (c), and with a patch of rod and cone loss (d); the legend applies to (b)–(d) only. The green dashed vertical lines mark the boundaries of rod (and cone) loss. To obtain $f_{init}(\theta)$ in (b)–(d), Eqs. (7) and (11) were solved using the finite difference method, with 4001 mesh points, where $p_r(\theta) = F(\theta)\bar{p}_r(\theta)$ and $p_c(\theta) = G(\theta)\bar{p}_c(\theta)$, and without treatment. (b) $F(\theta) = 1$ and $G(\theta) = 1$; (c) $F(\theta) = H(\theta_{r_1} - \theta) + H(\theta - \theta_{r_2})$ and $G(\theta) = 1$; (d) $F(\theta) = H(\theta_{r_1} - \theta) + H(\theta - \theta_{r_2})$ and $G(\theta) = H(\theta_{c_1} - \theta) + H(\theta - \theta_{c_2})$. Parameter values: $\xi = 0$; $(\theta_{r_1}, \theta_{r_2}) = (0.4, 0.6)$ in (c) and (d); $(\theta_{c_1}, \theta_{c_2}) = (0.4, 0.6)$ in (d). Remaining parameter values as in Table 2.

Table 1: Parameters employed in the dimensional model (Eqs. (1)–(6)). Parameters for which only the source is given are taken directly from the literature, those described as ‘calculated’ or ‘fitted’ are computed using data from the literature or from simulations, those described as ‘estimated’ are either estimated from relevant data in the literature or on the basis of biological reasoning, and those described as ‘chosen’ are free for us to choose. Values are given to three significant figures. See Supplementary Materials for justification of parameter values.

Parameter	Description	Value	Source
R	Retinal radial position	1.2×10^{-2} m	Oyster (1999)
Θ	Eccentricity of the ora serrata	1.33 rad	Curcio et al. (1990)
D_f	Trophic factor diffusivity	1.73×10^{-11} m ² s ⁻¹	Estimated using data from Jürgens et al. (1994)
α	Rate of trophic factor production by rods	1.81×10^{-17} Mm ² photoreceptors ⁻¹ s ⁻¹	Estimate using data from Eden et al. (2011)
ξ	Rate of trophic factor supply from treatment	0 to 1.72×10^{-6} Ms ⁻¹	Chosen
β	Rate of trophic factor consumption by cones	4.62×10^{-12} m ² photoreceptors ⁻¹ s ⁻¹	Estimate using data from Eden et al. (2011)
η	Rate of trophic factor decay	5.13×10^{-5} s ⁻¹	Estimated using data from Dörrbaum et al. (2018) and Eden et al. (2011)
ϕ_r	Rate of mutation-induced rod degeneration	2.10×10^{-8} s ⁻¹	Estimated using data from Curcio et al. (1993)
δ	Rate of trophic factor starvation-induced cone degeneration	2.87×10^{-7} s ⁻¹	Estimated using data from Kocaoglu et al. (2016)
μ_1	Growth rate of cone OS in phase 1	1.60×10^{-11} m s ⁻¹	Fitted using data from Guérin et al. (1993)
μ_2	Growth rate of cone OS in phase 2	3.07×10^{-12} m s ⁻¹	Fitted using data from Guérin et al. (1993)
δ_L	Rate of trophic factor starvation-induced cone OS degeneration	2.34×10^{-11} m s ⁻¹	Estimated using data from Kocaoglu et al. (2016)
\bar{p}_c^L	Healthy cone OS length	2.93×10^{-5} m	Kocaoglu et al. (2016)
f_{crit}	Trophic factor threshold concentration	3×10^{-9} to 3×10^{-5} M	Estimate
B_1	Cone profile parameter	1.73×10^{11} photoreceptors m ⁻²	Fitted using data from Curcio et al. (1990)
B_2	Cone profile parameter	1.76×10^{10} photoreceptors m ⁻²	Fitted using data from Curcio et al. (1990)
B_3	Rod profile parameter	8.84×10^{11} photoreceptors m ⁻² rad ⁻¹	Fitted using data from Curcio et al. (1990)
b_1	Cone profile parameter	54.1 rad ⁻¹	Fitted using data from Curcio et al. (1990)
b_2	Cone profile parameter	2.01 rad ⁻¹	Fitted using data from Curcio et al. (1990)
b_3	Rod profile parameter	2.31 rad ⁻¹	Fitted using data from Curcio et al. (1990)
\bar{f}_A	Mean trophic factor concentration	1×10^{-4} M	Calculated from simulations
\bar{p}_A	Mean photoreceptor density	1.11×10^{11} photoreceptors m ⁻²	Calculated using data from Curcio et al. (1990)
θ_{r_1}	Position of left-hand rod degenerate patch boundary	0 to Θ rad	Chosen
θ_{r_2}	Position of right-hand rod degenerate patch boundary	0 to Θ rad	Chosen
θ_{c_1}	Position of left-hand cone degenerate patch boundary	0 to Θ rad	Chosen
θ_{c_2}	Position of right-hand cone degenerate patch boundary	0 to Θ rad	Chosen
θ_{treat_1}	Position of left-hand limit of local treatment	0 to Θ rad	Chosen
θ_{treat_2}	Position of right-hand limit of local treatment	0 to Θ rad	Chosen
t_{crit}	Time at which treatment first applied	> 0 s	Chosen

2.1. Non-dimensionalisation

We recast the dimensional model in non-dimensional form to identify the dominant terms and simplify the equations. We scale the independent and dependent variables, and initial conditions as follows:

$$\theta = \Theta\theta^*, \quad t = \frac{t^*}{\delta}, \quad f = \tilde{f}_A f^*, \quad p_r = \tilde{p}_A p_r^*, \quad p_c = \tilde{p}_A p_c^*, \quad p_c^L = \tilde{p}_c^L p_c^{L*}, \quad f_{\text{init}} = \tilde{f}_A f_{\text{init}}^*.$$

Consequently, the scaled system is posed on the domain $\theta^* \in [0, 1]$. We rescale time by the rate of cone degeneration, δ , since this is the timescale of interest. The scaling factor \tilde{f}_A is the mean TF concentration under healthy conditions, found by taking the mean of the solution to Eqs. (1) and (5) at steady-state, with $p_r = \tilde{p}_r(\theta)$, $p_c = \tilde{p}_c(\theta)$ and $T(\theta) = 0$ (no treatment). The scaling factor \tilde{p}_A is the mean total photoreceptor density (rods and cones) under healthy conditions.

We define the following dimensionless parameters:

$$\begin{aligned} D_f^* &= \frac{D_f}{R^2 \Theta^2 \delta}, & \alpha^* &= \frac{\tilde{p}_A}{\delta \tilde{f}_A} \alpha, & \beta^* &= \frac{\tilde{p}_A}{\delta} \beta, & \eta^* &= \frac{\eta}{\delta}, & \phi_r^* &= \frac{\phi_r}{\delta}, & \mu_1^* &= \frac{\mu_1}{\delta \tilde{p}_c^L}, \\ \mu_2^* &= \frac{\mu_2}{\delta \tilde{p}_c^L}, & f_{\text{crit}}^* &= \frac{f_{\text{crit}}}{\tilde{f}_A}, & B_1^* &= \frac{B_1}{\tilde{p}_A}, & B_2^* &= \frac{B_2}{\tilde{p}_A}, & B_3^* &= \frac{\Theta}{\tilde{p}_A} B_3, & b_1^* &= \Theta b_1, \\ b_2^* &= \Theta b_2, & b_3^* &= \Theta b_3, & \theta_{r_1}^* &= \frac{\theta_{r_1}}{\Theta}, & \theta_{r_2}^* &= \frac{\theta_{r_2}}{\Theta}, & \theta_{c_1}^* &= \frac{\theta_{c_1}}{\Theta}, & \theta_{c_2}^* &= \frac{\theta_{c_2}}{\Theta}, \\ \xi^* &= \frac{\xi}{\delta \tilde{f}_A}, & \theta_{\text{treat}_1}^* &= \frac{\theta_{\text{treat}_1}}{\Theta}, & \theta_{\text{treat}_2}^* &= \frac{\theta_{\text{treat}_2}}{\Theta}, & t_{\text{crit}}^* &= \delta t_{\text{crit}}, & \delta_L^* &= \frac{\delta_L}{\delta \tilde{p}_c^L}, \end{aligned}$$

and note that $T(\theta, t) = T^*(\theta^*, t^*)$, where $T^*(\theta^*, t^*) = H(t^* - t_{\text{crit}}^*)$ or $(1 - [H(\theta_{\text{treat}_1}^* - \theta^*) + H(\theta^* - \theta_{\text{treat}_2}^*)])H(t^* - t_{\text{crit}}^*)$ (in the global and local treatment cases respectively); $C(\theta, f, p_c) = \tilde{p}_A \tilde{f}_A C^*(\theta^*, f^*, p_c^*)$, where $C^*(\theta^*, f^*, p_c^*) = p_c^* f^*$ or $p_{c_{\text{init}}}^*(\theta^*) f^*$ (in the without cone regeneration and with cone regeneration cases respectively); $\lambda_{1,2}(f) = \lambda_{1,2}^*(f^*)$, where $\lambda_1^*(f^*) = H(f^* - f_{\text{crit}}^*)$ and $\lambda_2^*(f^*) = 1 - H(f^* - f_{\text{crit}}^*)$; $\lambda_{3,4}(p_c^L) = \lambda_{3,4}^*(p_c^{L*})$, where $\lambda_3^*(p_c^L) = H(0.33 - p_c^{L*})$ and $\lambda_4^*(p_c^L) = 1 - H(0.33 - p_c^{L*})$; $\lambda_5(p_{c_{\text{init}}}^L) = \lambda_5^*(p_{c_{\text{init}}}^L)$, where $\lambda_5^*(p_{c_{\text{init}}}^L) = 1 - H(-p_{c_{\text{init}}}^L)$; $\lambda_6(p_c^L) = \lambda_6^*(p_c^{L*})$, where $\lambda_6^*(p_c^{L*}) = 1 - H(-p_c^{L*})$; $\tilde{p}_r(\theta) = \tilde{p}_A \tilde{p}_r^*(\theta^*)$, where $\tilde{p}_r^*(\theta^*) = B_3^* \theta^* e^{-b_3^* \theta^*}$; $\tilde{p}_c(\theta) = \tilde{p}_A \tilde{p}_c^*(\theta^*)$, where $\tilde{p}_c^*(\theta^*) = B_1^* e^{-b_1^* \theta^*} + B_2^* e^{-b_2^* \theta^*}$; $F(\theta) = F^*(\theta^*)$, where $F^*(\theta^*) = 1$ or $H(\theta_{r_1}^* - \theta^*) + H(\theta^* - \theta_{r_2}^*)$ (in the healthy and degenerate patch cases respectively); and $G(\theta) = G^*(\theta^*)$, where $G^*(\theta^*) = 1$ or $H(\theta_{c_1}^* - \theta^*) + H(\theta^* - \theta_{c_2}^*)$ (in the healthy and degenerate patch cases respectively).

Applying the above scalings to the dimensional governing equations, boundary and initial conditions (Eqs. (1)–(6)) and dropping the stars, we obtain the following dimensionless governing equations

$$\frac{\partial f}{\partial t} = \frac{D_f}{\sin(\Theta\theta)} \frac{\partial}{\partial \theta} \left(\sin(\Theta\theta) \frac{\partial f}{\partial \theta} \right) + \alpha p_r + \xi T(\theta, t) - \beta C(\theta, f, p_c) - \eta f, \quad (7)$$

$$\frac{\partial p_r}{\partial t} = \begin{cases} 0 & \text{w/o mutation-induced rod degeneration,} \\ -\phi_r p_r & \text{with mutation-induced rod degeneration,} \end{cases} \quad (8)$$

$$\frac{\partial p_c}{\partial t} = -p_c \lambda_2(f) \quad \text{w/o cone regeneration,} \quad (9)$$

$$\frac{\partial p_c^L}{\partial t} = \left[\mu_1 \lambda_3(p_c^L) + \mu_2 (1 - p_c^L) \lambda_4(p_c^L) \right] \lambda_1(f) \lambda_5(p_{c_{\text{init}}}) - \delta_L \lambda_2(f) \lambda_6(p_c^L) \quad \text{with cone regeneration,} \quad (10)$$

boundary conditions,

$$\frac{\partial f}{\partial \theta}(0, t) = 0 = \frac{\partial f}{\partial \theta}(1, t), \quad (11)$$

and initial conditions,

$$\begin{aligned} f(\theta, 0) &= f_{\text{init}}(\theta), & p_r(\theta, 0) &= p_{r_{\text{init}}}(\theta) = F(\theta) \tilde{p}_r(\theta), \\ p_c(\theta, 0) &= p_{c_{\text{init}}}(\theta) = G(\theta) \tilde{p}_c(\theta), & p_c^L(\theta, 0) &= p_{c_{\text{init}}}^L(\theta) = G(\theta), \end{aligned} \quad (12)$$

where $f_{\text{init}}(\theta)$ is the steady-state solution to Eqs. (7) and (11) with $p_r = p_{r_{\text{init}}}(\theta)$, $p_c = p_{c_{\text{init}}}(\theta)$ and $T(\theta) = 0$ (no treatment). See Table 2 for the dimensionless parameter values.

Table 2: Parameters employed in the non-dimensional model (Eqs. (7)–(12)). Values are given to three significant figures.

Parameter	Description	Value
D_f	Trophic factor diffusivity	0.237
α	Rate of trophic factor production by rods	7.01×10^4
ξ	Rate of trophic factor supply from treatment	0 to 6×10^4
β	Rate of trophic factor consumption by cones	1.79×10^6
η	Rate of trophic factor decay	1.79×10^2
ϕ_r	Rate of mutation-induced rod degeneration	7.33×10^{-2}
μ_1	Growth rate of cone OS in phase 1	1.90
μ_2	Growth rate of cone OS in phase 2	0.365
δ_L	Rate of trophic factor starvation-induced cone OS degeneration	2.79
f_{crit}	Trophic factor threshold concentration	3×10^{-5} or 0.3
B_1	Cone profile parameter	1.56
B_2	Cone profile parameter	0.158
B_3	Rod profile parameter	10.6
b_1	Cone profile parameter	71.8
b_2	Cone profile parameter	2.67
b_3	Rod profile parameter	3.06
θ_{r_1}	Position of left-hand rod degenerate patch boundary	0 to 1
θ_{r_2}	Position of right-hand rod degenerate patch boundary	0 to 1
θ_{c_1}	Position of left-hand cone degenerate patch boundary	0 to 1
θ_{c_2}	Position of right-hand cone degenerate patch boundary	0 to 1
θ_{treat_1}	Position of left-hand limit of local treatment	0 to 1
θ_{treat_2}	Position of right-hand limit of local treatment	0 to 1
t_{crit}	Time at which treatment first applied	> 0

3. Results: steady-state problem

In this section we consider the RdCVF Eqs. (7) and (11) at steady-state. Using a combination of asymptotic analysis and numerical simulations we seek to determine the spatial extent of a patch of cone loss, consequent upon the loss of a patch of rods, and the critical RdCVF treatment rate required to prevent cone degeneration following a patch of rod loss. The situation considered here mimics the early patchy loss of photoreceptors observed in RP patients (Cideciyan et al., 1998). In analysing the steady-state model we are making the simplifying assumptions that a patch of rod loss (or rod and cone loss) is already present, and neglecting mutation-induced rod loss. This allows for deeper analytical insight, allowing us to explore the effect of a patch of rod loss in isolation from mutation-induced rod loss. In Section 4, we drop these assumptions and consider the full dynamic problem.

3.1. Asymptotic analyses

There are a variety of scenarios to be considered:

- Rod loss only / rod and cone loss: either rods alone are removed from a patch, or rods and cones are removed from a patch, providing upper and lower bounds, respectively, on the predicted spatial extent of cone loss and critical treatment rate (see Section 3.2.1 for further details);

- Wide/narrow patch of photoreceptor loss: wide patches must be treated in an asymptotically distinct way from narrow patches (see Fig. A.1, Appendix A and Supplementary Materials), and represent late and early disease stages respectively;
- No treatment / treatment: in the untreated case we are interested in the extent of cone loss and in the treatment case we are interested in the critical RdCVF treatment rate required to prevent cone loss;
 - Global/local treatment: RdCVF treatment may be applied globally, across the whole retina, or locally, within the degenerate rod patch only. We are interested to see to what extent the critical treatment rate varies with global vs. local application.

We consider 8 steady-state (st-st) cases (of a possible 12):

1. St-st Case 1: wide patch of rod loss without treatment;
2. St-st Case 2: narrow patch of rod loss without treatment;
3. St-st Case 3: wide patch of rod and cone loss without treatment;
4. St-st Case 4: narrow patch of rod and cone loss without treatment;
5. St-st Case 5: wide patch of rod loss with global treatment;
6. St-st Case 6: wide patch of rod loss with local treatment;
7. St-st Case 7: narrow patch of rod loss with global treatment;
8. St-st Case 8: narrow patch of rod loss with local treatment.

We do not consider rod and cone loss with treatment (4 cases) since we are only interested in calculating the upper bound on the critical treatment rate. See Appendix A and the Supplementary Materials for details.

3.2. Numerical and analytical solutions

In this section we compare numerical solutions and analytical approximations to the steady-state problem in which rods (or rods and cones) have been lost from the interval $\theta \in (\theta_{r_1}, \theta_{r_2})$, with and without treatment. Analytical solutions provide deeper insight into the dominant mechanisms determining the behaviour of our models and are cheap to compute, while numerical solutions serve to confirm their accuracy and are more computationally expensive. To obtain numerical solutions we discretise Eqs. (7) and (11) at steady-state using a finite difference scheme and solve the resulting set of nonlinear algebraic equations for $f(\theta)$ using the Matlab routine `fsolve`, which employs a Trust–Region–Dogleg algorithm. We use between 401 and 4001 mesh points for all numerical solutions, using the maximum computationally feasible number of mesh points in each case. The upper bound of 4001 mesh points is chosen such that the distance between mesh points corresponds to the average width of a photoreceptor. Analytical approximations are derived in Appendix A and the Supplementary Materials. We consider the 8 cases described in Section 3.1, dividing each of these cases into the two subcases described below.

Given that the foveal cone density is high and the rod density low (and hence the foveal rod:cone ratio is low) compared to the rest of the retina, the RdCVF supply to each cone is far lower in the fovea than in the rest of the retina. This suggests that either there is a surplus of RdCVF produced outside of the fovea (Subcase (i)), or that the cones in the fovea require less RdCVF than elsewhere (Subcase (ii)). Since the RdCVF threshold has not been measured, it is of value to consider both subcases here:

1. Subcase (i): low (homogeneous) RdCVF threshold concentration $f_{\text{crit}} = 3 \times 10^{-5}$: chosen such that $f_{\text{crit}} < f(\theta)$ for all $\theta \in [0, 1]$ in the healthy retina (where $p_r(\theta) = \tilde{p}_r(\theta)$ and $p_c(\theta) = \tilde{p}_c(\theta)$);
2. Subcase (ii): high (heterogeneous) RdCVF threshold concentration $f_{\text{crit}} = f_{\text{crit}}(\theta) = f_{\text{crit}_1}H(0.13 - \theta) + f_{\text{crit}_2}(1 - H(0.13 - \theta))$: where $f_{\text{crit}_1} = 3 \times 10^{-5}$, $f_{\text{crit}_2} = 0.3$ and the switch point, $\theta = 0.13$, are chosen such that $f_{\text{crit}}(\theta) < f(\theta)$ for all $\theta \in [0, 1]$ in the healthy retina.

In Subcase (i) (low f_{crit}), the threshold is spatially uniform and chosen to be low enough such that cones will not degenerate in the healthy retina. In Subcase (ii) (high f_{crit}), we assume that the cone-rich fovea is afforded special protection against low RdCVF levels, such that f_{crit} is lower in that region ($\theta \leq 0.13$) than in the rest of the retina. The existence and nature of this special protection remains an open question; however, such a situation is not without precedent given the maintained upregulation of basic fibroblast growth factor and glial fibrillary acidic protein expression measured at the edge of the mouse retina to protect against stress in that region (Mervin and Stone, 2002). For notational simplicity, we shall refer to the high f_{crit} subcase simply as $f_{\text{crit}} = 0.3$ in what follows. Note that throughout this section, we consider only the region $\theta \in (0.13, 1]$ in the high f_{crit} subcase.

3.2.1. No treatment

We begin by considering the untreated cases (St-st Cases 1–4), in which $\xi = 0$. Here we are interested in predicting the interval over which cones will degenerate, $(\theta_{\text{crit}_1}, \theta_{\text{crit}_2})$, following the removal of a patch of rods from the interval $\theta \in (\theta_{r_1}, \theta_{r_2})$, and the conditions under which cone degeneration will expand beyond the rod degenerate patch. The boundaries of the resultant patch of cone loss, $(\theta_{\text{crit}_1}, \theta_{\text{crit}_2})$, are calculated as the positions at which $f(\theta) = f_{\text{crit}}$ (between which $f(\theta) < f_{\text{crit}}$) local to the patch of rod loss. In the full dynamic problem (see Section 4.1), $f(\theta)$ rises as cones degenerate, shrinking the interval over which $f(\theta) < f_{\text{crit}}$. Here, we derive upper and lower bounds on the cone degenerate patch width. The upper bound is found in the case where only rods are removed (St-st Cases 1 and 2), such that cones continue to consume RdCVF within the degenerate patch, expanding the interval over which $f(\theta) < f_{\text{crit}}$. The lower bound is found in the case where both rods and cones are removed from $\theta \in (\theta_{r_1}, \theta_{r_2})$ (St-st Cases 3 and 4, where $\theta_{c_1} = \theta_{r_1}$ and $\theta_{c_2} = \theta_{r_2}$), such that cones do not consume RdCVF within the degenerate patch, reducing the interval over which $f(\theta) < f_{\text{crit}}$. We note that, in calculating the lower bound this way, we assume that cones do not degenerate beyond the rod degenerate patch. Where cone loss does exceed the rod patch, this method acts as a confirmation that this will occur, rather than as a lower bound.

Fig. S2 (Supplementary Material) shows analytical solutions for the distance between the left-hand rod and cone degenerate patch boundaries, $\theta_{r_1} - \theta_{\text{crit}_1}$, as a function of θ_{r_1} , and similarly for the distance between the right-hand rod and cone degenerate patch boundaries, $\theta_{\text{crit}_2} - \theta_{r_2}$, as a function of θ_{r_2} . We consider St-st Case 1 (rod loss only — (a) and (b)), and St-st Case 3 (rod and cone loss — (c) and (d)), corresponding to the upper and lower bounds on cone degenerate patch width, respectively. For each of these cases, we consider the subcases (ii) $f_{\text{crit}} = 0.3$ ((a) and (c)) and (i) $f_{\text{crit}} = 3 \times 10^{-5}$ ((b) and (d)).

The equations plotted in Fig. S2 ($\theta_{r_1} - \theta_{\text{crit}_1}$ and $\theta_{\text{crit}_2} - \theta_{r_2}$, where θ_{crit_1} and θ_{crit_2} are as given in Eqs. (A.22) and (A.23) in (a) and (b), and Eqs. (S.21)–(S.24) (Supplementary Material) in (c) and (d)) are all monotone decreasing functions of θ_{r_1} and θ_{r_2} . However, θ_{crit_1} and θ_{crit_2} cannot in practice go outside of the retina and hence cannot exceed the interval $\theta \in [0, 1]$. Therefore, we set $\theta_{\text{crit}_1} = 1$ where $\theta_{\text{crit}_1} > 1$ and $\theta_{\text{crit}_2} = 0$ where $\theta_{\text{crit}_2} < 0$. This results in upward jumps on the right-hand side of the domain in $\theta_{r_1} - \theta_{\text{crit}_1}$ in (a)–(d) and an upward jump on the left-hand side of the domain in $\theta_{\text{crit}_2} - \theta_{r_2}$ in (d). Apart from these jumps, the curves for $\theta_{r_1} - \theta_{\text{crit}_1}$ and $\theta_{\text{crit}_2} - \theta_{r_2}$ are identical. Note that, for the wide patch case, the predicted position of one cone degenerate patch boundary does not depend upon the other cone degenerate patch boundary (unlike in the narrow patch case), since the boundary layers surrounding the rod degenerate patch boundaries are separated by a central outer region, such that they do not ‘interact’. The maximum spatial extent of cone loss remains within the boundaries of rod loss in (b) and (d), but may exceed it close to the fovea (centred at $\theta = 0$) in (a) and (c).

Fig. 3 shows numerical and analytical bounds on cone degenerate patch size for a wide rod degenerate patch (St-st Cases 1 and 3). The cone degenerate patch width, $\theta_{\text{crit}_2} - \theta_{\text{crit}_1}$, increases monotonically with increasing rod degenerate patch width, $\theta_{r_2} - \theta_{r_1}$, and with decreasing rod degenerate patch centre position,

$(\theta_{r_1} + \theta_{r_2})/2$. For narrower rod loss patches in (c), and especially (d), the predicted lower bound on cone degenerate patch width is zero. The difference between rod and cone degenerate patch widths, $(\theta_{\text{crit}_2} - \theta_{\text{crit}_1}) - (\theta_{r_2} - \theta_{r_1})$, is a monotone (not strictly) decreasing function of rod degenerate patch centre position. The cone degenerate patch width is less than the rod degenerate patch width except in (a) (rod loss only with $f_{\text{crit}} = 0.3$), where it may exceed rod degenerate patch width close to the fovea, doing so for a wider range of patch positions for narrower rod loss patches. The predicted cone degenerate patch width is lower for the rod and cone loss case ((a) and (b), lower bound) than for the rod loss only case ((c) and (d), upper bound) as would be expected. Numerical and analytical results match closely, except for the width 0.1 rod and cone degenerate patch case in (c), where the wide patch assumption breaks down, since $0.1 \sim O(\epsilon^{1/2})$.

Fig. 4 shows numerical and analytical bounds on cone degenerate patch size for a narrow rod degenerate patch (St-st Cases 2 and 4). We plot only the $f_{\text{crit}} = 0.3$ subcase here since, for the $f_{\text{crit}} = 3 \times 10^{-5}$ subcase, the predicted cone loss patch width is always zero. As in Fig. 3, cone degenerate patch width increases monotonically with increasing rod degenerate patch width and with decreasing rod degenerate patch centre position, while the difference between rod and cone degenerate patch widths is a monotone (not strictly) decreasing function of rod degenerate patch centre position. The cone degenerate patch width may exceed the rod degenerate patch width close to the fovea in (a). Numerical and analytical solutions agree well in (a) and on the right-hand side in (b); however, the scaling assumptions break down close to the fovea, such that the analytical solution predicts cone loss on the left-hand side in (b), where the numerical solution remains zero.

3.2.2. Treatment

In this section we compare numerical and analytical estimates for the critical treatment rate, ξ_{crit} . This is the minimum RdCVF treatment rate required to prevent cone degeneration in a given scenario. We consider just the rod loss only case, since this provides an upper bound on ξ_{crit} , which is of greater clinical relevance, given that this would theoretically guarantee the avoidance of cone degeneration under the trophic factor hypothesis. We also consider analytical estimates for the eccentricity of the minimum TF concentration local to the rod degenerate patch, θ_{crit} , when $\xi = \xi_{\text{crit}}$. This is the point at which cone loss will initiate if ξ drops below ξ_{crit} . Numerical solutions are obtained using the Matlab routine `fminsearch` (which uses a simplex search method) to find the value of ξ at which the minimum value of $f(\theta)$ local to the rod degenerate patch is equal to f_{crit} . This involves solving Eqs. (7) and (11) for $f(\theta)$ at steady-state using `fsolve` at each iteration. Analytical solutions are obtained by employing the Matlab routine `fsolve` to solve the implicit equations (S.46) and (S.47) for a wide patch of rod loss with global treatment, (S.59) and (S.60) for a wide patch of rod loss with local treatment, (S.71) and (S.72) for a narrow patch of rod loss with global treatment, and (S.83) and (S.84) for a narrow patch of rod loss with local treatment.

Fig. 5 shows numerical and analytical estimates of the critical treatment rate for a wide rod degenerate patch (St-st Cases 5 and 6). The critical treatment rate increases monotonically with decreasing rod degenerate patch centre position and left boundary position in all cases. The critical treatment rate depends almost exclusively upon the rod degenerate patch left boundary position, rather than the rod degenerate patch width or right boundary position. The critical treatment rate is $O(10^4)$ for $f_{\text{crit}} = 0.3$ ((a) and (b)) and $O(1)$ for $f_{\text{crit}} = 3 \times 10^{-5}$ ((c) and (d)), a lower rate being sufficient to raise the TF concentration above the lower critical threshold. Numerical and analytical solutions agree well.

Fig. 6 shows numerical and analytical estimates of the critical treatment rate for a narrow rod degenerate patch (St-st Cases 7 and 8). We consider only the $f_{\text{crit}} = 0.3$ subcase here since there is no cone loss for $f_{\text{crit}} = 3 \times 10^{-5}$ and hence no treatment is required in this subcase. As with the wide rod degenerate patch case, the critical treatment rate increases monotonically with decreasing rod degenerate patch centre position and left boundary position in all cases. Unlike in the wide rod degenerate patch case, the critical treatment

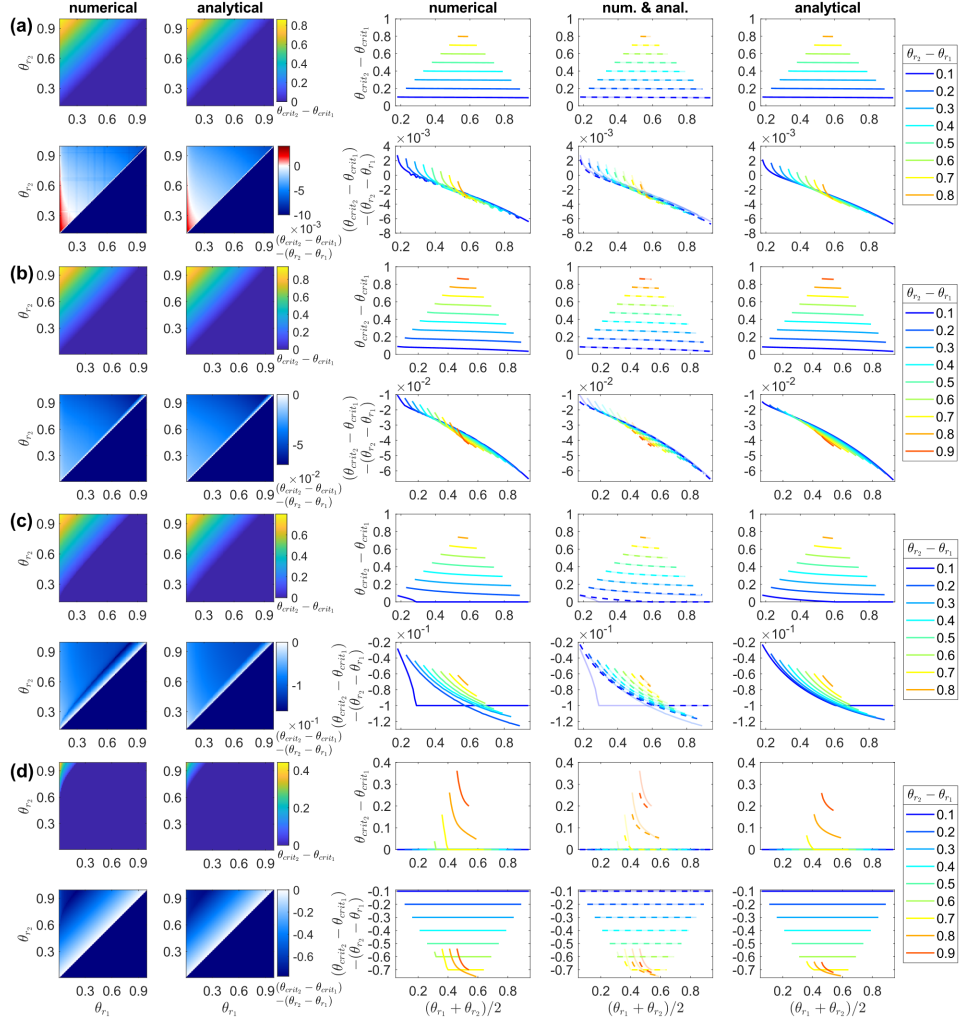


Figure 3: Numerical and analytical bounds on cone degenerate patch size using the steady-state model — wide rod degenerate patch (St-st Cases 1 and 3). (a)–(d) first row: maximum/minimum cone degenerate patch width, $\theta_{\text{crit}2} - \theta_{\text{crit}1}$; second row: difference between rod and maximum/minimum cone degenerate patch widths, $(\theta_{\text{crit}2} - \theta_{\text{crit}1}) - (\theta_{r2} - \theta_{r1})$; columns 1 and 2: variation of patch widths and width differences in $(\theta_{r1}, \theta_{r2})$ parameter space; columns 3–5: variation of patch widths and width differences with rod degenerate patch centre position, $(\theta_{r1} + \theta_{r2})/2$, each curve representing a constant rod degenerate patch width, $\theta_{r2} - \theta_{r1}$; columns 1 and 3: numerical solutions; columns 2 and 5: analytical approximations; column 4: numerical (solid curves) and analytical (dashed curves) solutions compared. (a) St-st Case 1(ii): rod loss only with $f_{\text{crit}} = 0.3$; (b) St-st Case 1(i): rod loss only with $f_{\text{crit}} = 3 \times 10^{-5}$; (c) St-st Case 3(ii): rod and cone loss with $f_{\text{crit}} = 0.3$; (d) St-st Case 3(i): rod and cone loss with $f_{\text{crit}} = 3 \times 10^{-5}$. (a) and (b) upper bounds on cone degenerate patch widths; (c) and (d) lower bounds on cone degenerate patch widths. Cone degenerate patch width increases monotonically with increasing rod degenerate patch width and with decreasing rod degenerate patch centre position. Cone degenerate patch width is less than rod degenerate patch width except in (a), where it may exceed rod degenerate patch width close to the fovea (centred at $\theta = 0$). Numerical solutions were obtained by calculating the eccentricities at which $f(\theta) = f_{\text{crit}}$, solving Eqs. (7) and (11) for $f(\theta)$ at steady-state using the finite difference method, with 4001 mesh points, where $p_r(\theta) = F(\theta)\bar{p}_r(\theta)$, $p_c(\theta) = G(\theta)\bar{p}_c(\theta)$, $F(\theta) = H(\theta_{r1} - \theta) + H(\theta - \theta_{r2})$, $G(\theta) = 1$ in (a) and (b), and $G(\theta) = H(\theta_{c1} - \theta) + H(\theta - \theta_{c2})$, with $\theta_{c1} = \theta_{r1}$ and $\theta_{c2} = \theta_{r2}$, in (c) and (d), and without treatment or cone regeneration. Analytical solutions are plotted using Eqs. (A.22) and (A.23) in (a) and (b), and using Eqs. (S.21)–(S.24) in (c) and (d). Parameter values: $\xi = 0$ and $\epsilon = 10^{-2}$. Remaining parameter values as in Table 2.

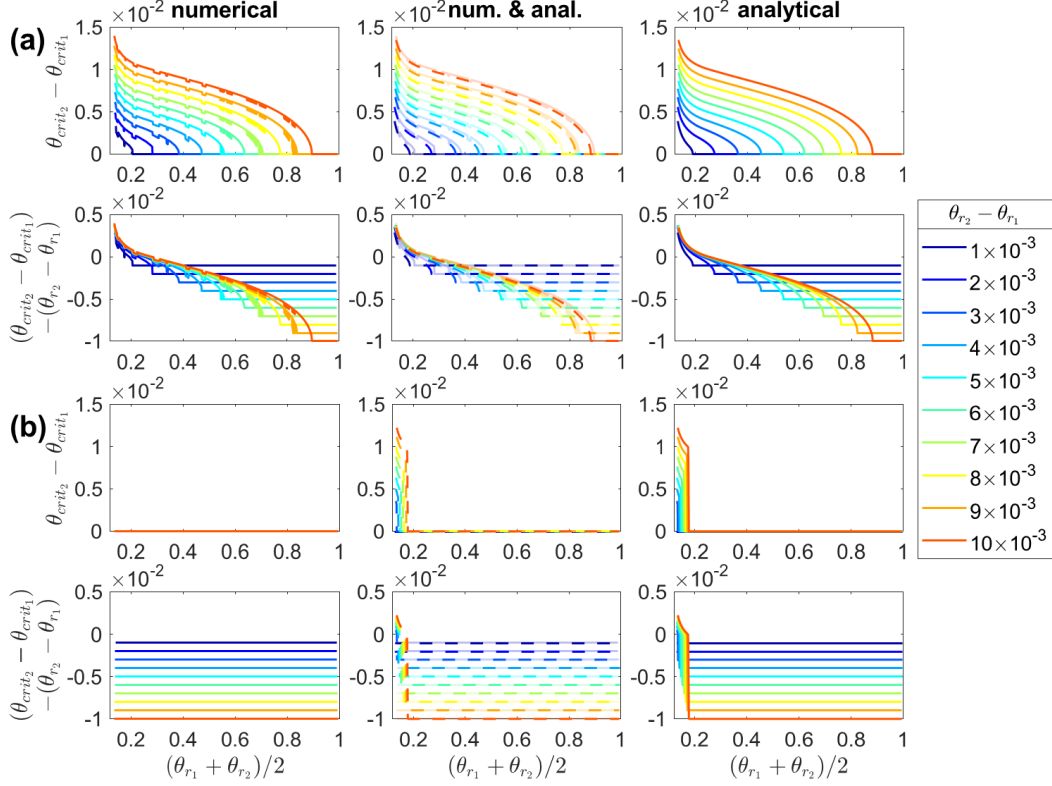


Figure 4: Numerical and analytical bounds on cone degenerate patch size using the steady-state model — narrow rod degenerate patch (St-st Cases 2 and 4). Graphs show variation of maximum/minimum cone degenerate patch widths, and differences between rod and maximum/minimum cone degenerate patch widths with rod degenerate patch centre position, $(\theta_{r1} + \theta_{r2})/2$, each curve representing a constant rod degenerate patch width, $\theta_{r2} - \theta_{r1}$. (a) and (b) first row: maximum/minimum cone degenerate patch width, $\theta_{crit2} - \theta_{crit1}$; second row: difference between rod and maximum/minimum cone degenerate patch widths, $(\theta_{crit2} - \theta_{crit1}) - (\theta_{r2} - \theta_{r1})$; column 1: numerical solutions; column 3: analytical approximations; column 2: numerical (solid curves) and analytical (dashed curves) solutions compared. (a) St-st Case 2(ii) (rod loss only): upper bounds on cone degenerate patch widths; (b) St-st Case 4(ii) (rod and cone loss): lower bounds on cone degenerate patch widths. Cone degenerate patch width increases monotonically with increasing rod degenerate patch width and with decreasing rod degenerate patch centre position. Cone degenerate patch width may exceed rod degenerate patch width close to the fovea (centred at $\theta = 0$). Numerical solutions were obtained by calculating the eccentricities at which $f(\theta) = f_{crit}$, solving Eqs. (7) and (11) for $f(\theta)$ at steady-state using the finite difference method, with 4001 mesh points, where $p_r(\theta) = F(\theta)\tilde{p}_r(\theta)$, $p_c(\theta) = G(\theta)\tilde{p}_c(\theta)$, $F(\theta) = H(\theta_{r1} - \theta) + H(\theta - \theta_{r2})$, $G(\theta) = 1$ in (a), and $G(\theta) = H(\theta_{c1} - \theta) + H(\theta - \theta_{c2})$, with $\theta_{c1} = \theta_{r1}$ and $\theta_{c2} = \theta_{r2}$, in (b), and without treatment or cone regeneration. Analytical solutions are plotted using Eqs. (S.7)–(S.10) in (a), and using Eqs. (S.31)–(S.34) in (b). Parameter values: $f_{crit} = 0.3$, $\xi = 0$ and $\epsilon = 10^{-2}$. Remaining parameter values as in Table 2.

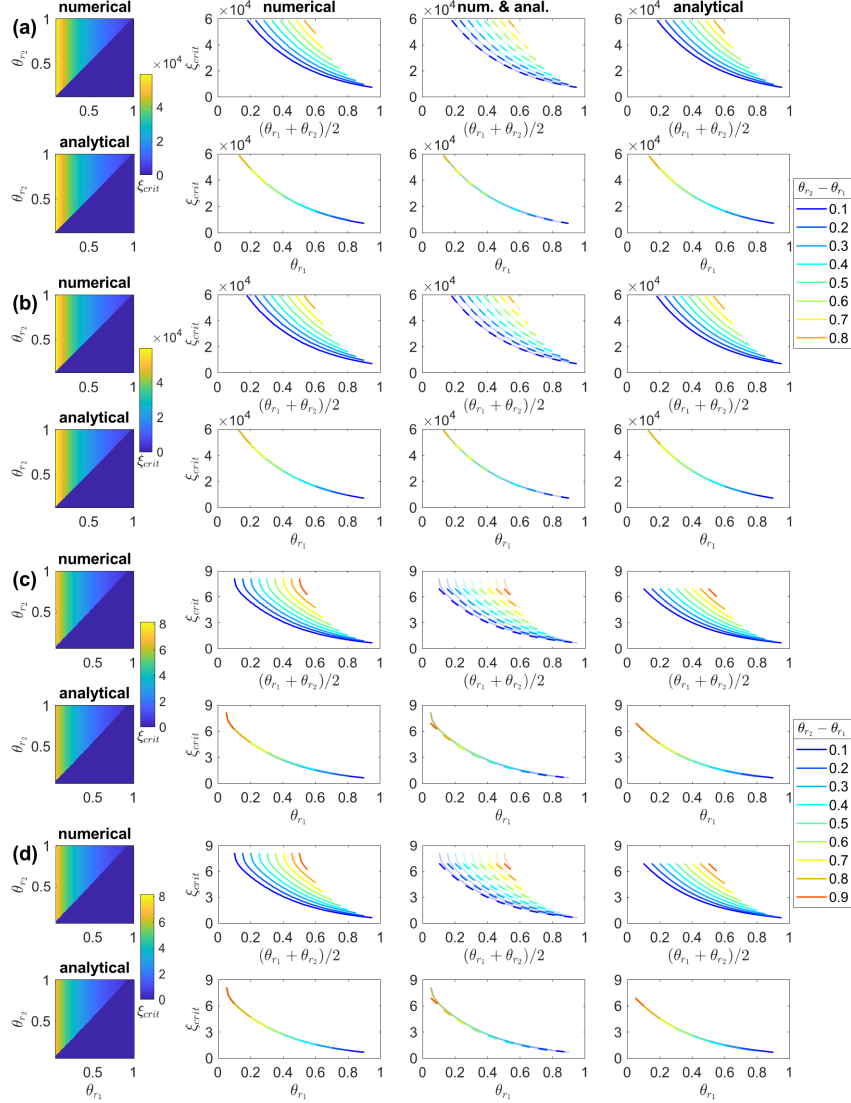


Figure 5: Numerical and analytical estimates of the critical treatment rate using the steady-state model — wide rod degenerate patch with rod loss only (St-st Cases 5 and 6). All plots show the critical treatment rate, ξ_{crit} . (a) and (c) St-st Case 5 (global treatment), (b) and (d) St-st Case 6 (local treatment); (a) and (b) Subcase (ii) ($f_{\text{crit}} = 0.3$); (c) and (d) Subcase (i) ($f_{\text{crit}} = 3 \times 10^{-5}$). (a)–(d) column 1: numerical (top) and analytical (bottom) estimates of the variation in the critical treatment rate over $(\theta_{r_1}, \theta_{r_2})$ parameter space; columns 2–4: variation in the critical treatment rate with rod degenerate patch centre position, $(\theta_{r_1} + \theta_{r_2})/2$ (top), and with rod degenerate patch left boundary position, θ_{r_1} (bottom), each curve representing a constant rod degenerate patch width, $\theta_{r_2} - \theta_{r_1}$; column 2: numerical solutions; column 4: analytical approximations; column 3: numerical (solid curves) and analytical (dashed curves) solutions compared. The critical treatment rate increases monotonically with decreasing rod degenerate patch centre/left boundary position in all cases. The critical treatment rate depends almost entirely upon the rod degenerate patch left boundary position. Numerical solutions were obtained by using the Matlab routine `fminsearch` to calculate the critical treatment rate at which $\min(f(\theta)) = f_{\text{crit}}$ within $\theta \in [\theta_{r_1}, \theta_{r_2}]$, solving Eqs. (7) and (11) for $f(\theta)$ at steady-state at each iteration using the finite difference method, with 401 mesh points, where $p_r(\theta) = F(\theta)\tilde{p}_r(\theta)$, $p_c(\theta) = G(\theta)\tilde{p}_c(\theta)$, $F(\theta) = H(\theta_{r_1} - \theta) + H(\theta - \theta_{r_2})$, $G(\theta) = 1$, $T(\theta) = 1$ for (a) and (c), and $T(\theta) = 1 - F(\theta)$ for (b) and (d). Analytical solutions are obtained by implicitly solving Eqs. (S.46) and (S.47) in (a) and (c), and Eqs. (S.59) and (S.60) in (b) and (d). The parameter $\epsilon = 10^{-2}$. Remaining parameter values as in Table 2.

rate depends upon both the rod degenerate patch left and right boundary positions, and hence upon the patch width (compare Fig. 5), increasing monotonically with increasing rod degenerate patch width. Numerical and analytical solutions match closely.

Fig. 7 compares analytical estimates of the critical treatment rate for the global and local treatment cases (St-st Cases 5–8). The critical treatment rate is almost identical for global and local treatment in the wide patch case (St-st Cases 5 and 6), for both $f_{\text{crit}} = 0.3$ (a) and $f_{\text{crit}} = 3 \times 10^{-5}$ (b), while the critical treatment rate is higher for local than for global treatment for the narrowest patches close to the fovea (centred at $\theta = 0$) in the narrow patch case (St-st Cases 7 and 8, with $f_{\text{crit}} = 0.3$) (c).

Fig. S3 shows analytical estimates of the normalised eccentricity of the minimum TF concentration, $\hat{\theta}_{\text{crit}} = (\theta_{\text{crit}} - \theta_{r_1})/(\theta_{r_2} - \theta_{r_1})$, for both wide and narrow rod degenerate patches (St-st Cases 5–8). The variable $\hat{\theta}_{\text{crit}}$ increases monotonically with decreasing rod degenerate patch width, and with increasing rod degenerate patch centre position and left boundary position in all cases. Further, $\hat{\theta}_{\text{crit}}$ remains close to 0 for patches of width $\theta_{r_2} - \theta_{r_1} \geq 0.2$ ((a)–(d)) and close to 0.5 for narrow patches ((e) and (f)).

4. Results: dynamic problem

Having explored the steady-state problem, we now consider the full dynamic problem, consisting of Eqs. (7)–(9) and (11)–(12) in the no cone regeneration/no treatment case, and Eqs. (7), (8) and (10)–(12) in the cone regeneration/treatment case. In both cases, equations are solved using the method of lines (as for a similar system of equations in one of our earlier studies; Roberts et al., 2017). This involves discretising in space and then integrating in time using the Matlab routine `ode15s`, a variable-step, variable-order solver, chosen since this is a stiff problem, involving multiple timescales. We used either the default relative and absolute error tolerances (10^{-3} and 10^{-6} respectively) or more stringent tolerances (10^{-6} and 10^{-10} respectively) in all simulations. As in Section 3.2, we use between 401 and 4001 mesh points for all simulations. The initial TF profile, $f(\theta, 0) = f_{\text{init}}(\theta)$, is calculated by discretising Eqs. (7) and (11) at steady-state and solving the consequent system of nonlinear algebraic equations using the Matlab routine `fsolve` with $p_r = p_{r_{\text{init}}}(\theta)$ and $p_c = p_{c_{\text{init}}}(\theta)$.

There are a variety of scenarios to be considered:

- No rod loss patch / rod loss patch: either the retina is healthy initially or there is a single patch of rod loss;
 - Wide/narrow patch: representing late and early disease stages respectively;
 - Patch eccentricity: we will consider patches on the left-hand side (parafovea/perifovea), centre (mid-periphery) and right-hand side (far-periphery) of the domain;
- No mutation-induced rod loss / mutation-induced rod loss: in the absence of mutation-induced rod loss, photoreceptor (cone) degeneration is due solely to RdCVF starvation, whereas, in the presence of mutation-induced rod loss, rods also degenerate exponentially across the domain over time;
- No treatment / treatment: we consider both the natural evolution of retinal degeneration in the absence of treatment, and the effect of RdCVF treatment upon cone degeneration and cone OS regeneration;
 - Global/local treatment: as in Section 3, RdCVF treatment may be applied globally, across the whole retina, or locally, within a degenerate rod patch only.

Unlike in Section 3, we do not consider the scenario in which both rods and cones have been lost from a patch initially, since, as demonstrated in that section, cone loss seldom proceeds to exceed rod loss.

We consider the following 5 dynamic (dyn) cases (/combinations of scenarios):

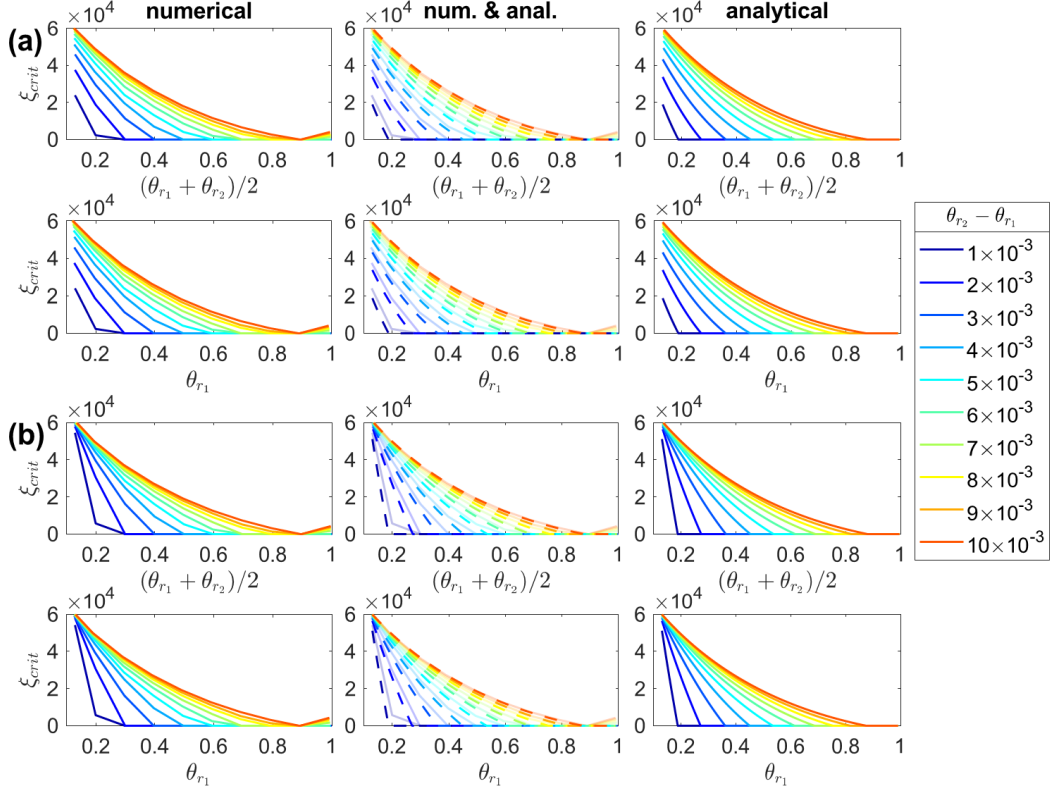


Figure 6: Numerical and analytical estimates of the critical treatment rate using the steady-state model — narrow rod degenerate patch with rod loss only (St-st Cases 7 and 8). All plots show the critical treatment rate, ξ_{crit} . (a) St-st Case 7(ii) (global treatment), (b) St-st Case 8(ii) (local treatment). (a) and (b) variation in the critical treatment rate with rod degenerate patch centre position, $(\theta_{r_1} + \theta_{r_2})/2$ (top), and with rod degenerate patch left boundary position, θ_{r_1} (bottom), each curve representing a constant rod degenerate patch width, $\theta_{r_2} - \theta_{r_1}$; column 1: numerical solutions; column 2: numerical (solid curves) and analytical (dashed curves) solutions compared. The critical treatment rate depends upon both the rod degenerate patch left and right boundary positions, and hence the patch width (compare Fig. 5). The critical treatment rate increases monotonically with increasing rod degenerate patch width and with decreasing patch centre/left boundary position in all cases. Numerical solutions were obtained by using the Matlab routine `fminsearch` to calculate the critical treatment rate at which $\min(f(\theta)) = f_{crit}$ within $\theta \in [\theta_{r_1}, \theta_{r_2}]$, solving Eqs. (7) and (11) for $f(\theta)$ at steady-state at each iteration using the finite difference method, with 1001 mesh points, where $p_r(\theta) = F(\theta)\bar{p}_r(\theta)$, $p_c(\theta) = G(\theta)\bar{p}_c(\theta)$, $F(\theta) = H(\theta_{r_1} - \theta) + H(\theta - \theta_{r_2})$, $G(\theta) = 1$, $T(\theta) = 1$ for (a) and $T(\theta) = 1 - F(\theta)$ for (b). Analytical solutions are obtained by implicitly solving Eqs. (S.71) and (S.72) in (a) and Eqs. (S.83) and (S.84) in (b). Parameter values: $f_{crit} = 0.3$ and $\epsilon = 10^{-2}$. Remaining parameter values as in Table 2.

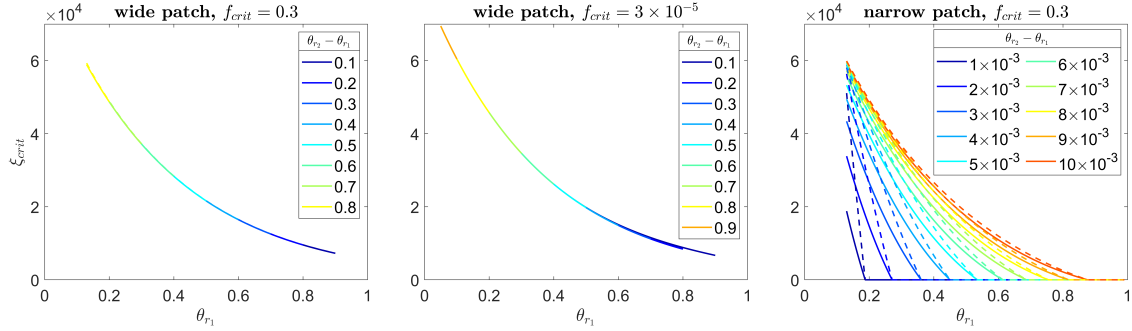


Figure 7: Comparison of critical treatment rates under global and local treatment using the steady-state model — wide and narrow rod degenerate patch (St-st Cases 5–8). All plots show analytical estimates of the variation in critical treatment rate, ξ_{crit} , with rod degenerate patch left boundary position, θ_{r_1} , using the steady-state model. Each curve represents a constant rod degenerate patch width, $\theta_{r_2} - \theta_{r_1}$. Solid curves: global treatment; dashed curves: local treatment. (a) St-st Cases 5(ii) and 6(ii): wide rod degenerate patch with $f_{\text{crit}} = 0.3$; (b) St-st Cases 5(i) and 6(i): wide patch with $f_{\text{crit}} = 3 \times 10^{-5}$; (c) St-st Cases 7(ii) and 8(ii): narrow patch with $f_{\text{crit}} = 0.3$. The critical treatment rate is almost identical for global and local treatment in the wide patch case, while the critical treatment rate is higher for local than for global treatment for the narrowest patches close to the fovea (centred at $\theta = 0$) in the narrow patch case. Analytical solutions and parameter values are identical to those for the corresponding cases in Figs. 5 and 6.

1. Dyn Case 1: wide and narrow patches of rod loss, at left/central/right positions, without mutation-induced rod loss or treatment;
2. Dyn Case 2: mutation-induced rod loss without patch loss or treatment;
3. Dyn Case 3: wide and narrow patches of rod loss, at left/central/right positions, with mutation-induced rod loss and without treatment;
4. Dyn Case 4: wide and narrow patches of rod loss, at left-hand positions, with global and local treatment, and without mutation-induced rod loss;
5. Dyn Case 5: mutation-induced rod loss with global treatment and without patch loss.

As in Section 3, we also consider the subcases (i) $f_{\text{crit}} = 3 \times 10^{-5}$ and (ii) $f_{\text{crit}} = 0.3$. Unlike in Section 3, we now solve and plot across the whole domain for the $f_{\text{crit}} = 0.3$ subcase.

4.1. No treatment

Fig. 8 shows RdCVF and photoreceptor dynamics following the complete removal of rods from the interval $(\theta_{r_1}, \theta_{r_2})$ (Dyn Case 1). Cones degenerate within all wide patches of rod loss in (b) ($f_{\text{crit}} = 3 \times 10^{-5}$) and (c) ($f_{\text{crit}} = 0.3$), and within all narrow patches of rod loss in (c), but only in the left-hand narrow patch of rod loss in (b). The boundaries of cone loss remain within those of rod loss for all patches in (b) and for central and right patches in (c); however, cone loss exceeds rod loss on the left-hand side of the left wide patch and on both the left- and right-hand sides of the left narrow patch in (c). This agrees nicely with our predictions in Figs. 3 and 4.

Fig. 9 shows RdCVF and photoreceptor dynamics with mutation-induced rod degeneration (Dyn Case 2). Cone degeneration initiates at the fovea ($\theta = 0$) in (a) ($f_{\text{crit}} = 3 \times 10^{-5}$) and at $\theta = 0.13$ in (b) ($f_{\text{crit}} = 0.3$), spreading peripherally (rightwards) in both cases. The retina is mostly preserved for $\theta \leq 0.13$ in (b) due to the lower value of $f_{\text{crit}} = f_{\text{crit}_1} = 3 \times 10^{-5}$ in that region (see discussion in Section 3.2 for more details); however, given enough time, photoreceptors will also degenerate in this region, as in (a). The rate at which cone degeneration spreads across the domain increases over time as it propagates peripherally.

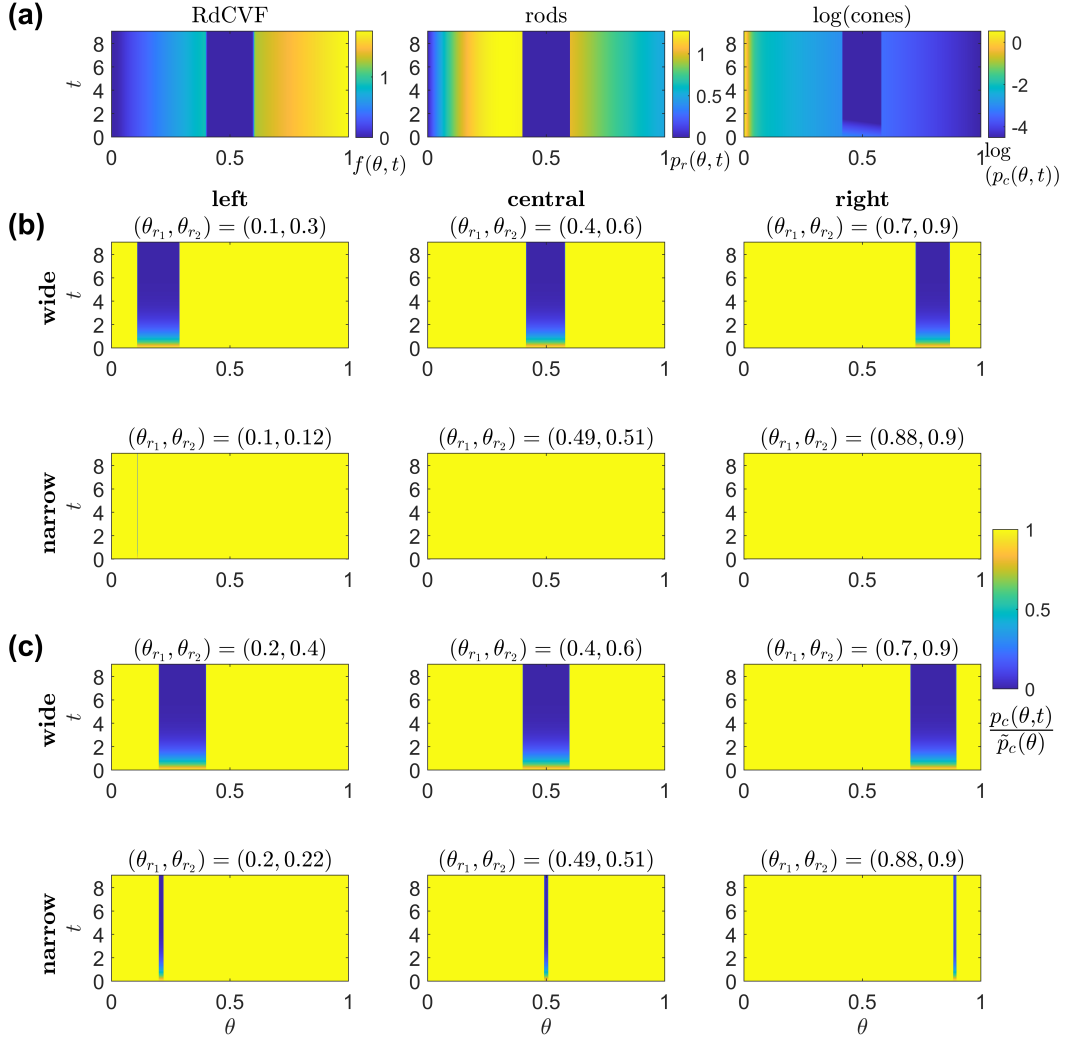


Figure 8: Simulations of RdCVF and photoreceptor dynamics following the complete removal of rods from the interval $(\theta_{r_1}, \theta_{r_2})$ — dynamic model (Dyn Case 1). (a) left-to-right: TF concentration, $f(\theta, t)$, rod density, $p_r(\theta, t)$, and the natural logarithm of the cone density, $\log(p_c(\theta, t))$ (colour scale lower threshold: $\log(\min(p_{c_{\text{init}}(\theta)))$), for Dyn Case 1(i): $f_{\text{crit}} = 3 \times 10^{-5}$ and $(\theta_{r_1}, \theta_{r_2}) = (0.4, 0.6)$ (the same as for the top-middle panel in (b)). (b) proportional cone loss, $p_c(\theta, t)/\bar{p}_c(\theta)$, for Dyn Case 1(i): $f_{\text{crit}} = 3 \times 10^{-5}$. (c) proportional cone loss for Dyn Case 1(ii): $f_{\text{crit}} = 0.3$. Rods are removed from wide (width $O(1)$) and narrow (width $O(\epsilon)$) intervals, on the left-hand side (parafovea/perifovea), centre (mid-periphery) and right-hand side (far-periphery) of the domain. All simulations span the period of one year in dimensional variables. Cones degenerate within all wide patches of rod loss ((b) and (c)), and within all narrow patches of rod loss in (c), but only in the left-hand narrow patch of rod loss in (b). Eqs. (7)–(9) and (11)–(12) were solved using the method of lines, with 4001 mesh points, $F(\theta) = H(\theta_{r_1} - \theta) + H(\theta - \theta_{r_2})$ and $G(\theta) = 1$, and without treatment, cone regeneration or mutation-induced rod degeneration. The parameter $\xi = 0$. Remaining parameter values as in Table 2.

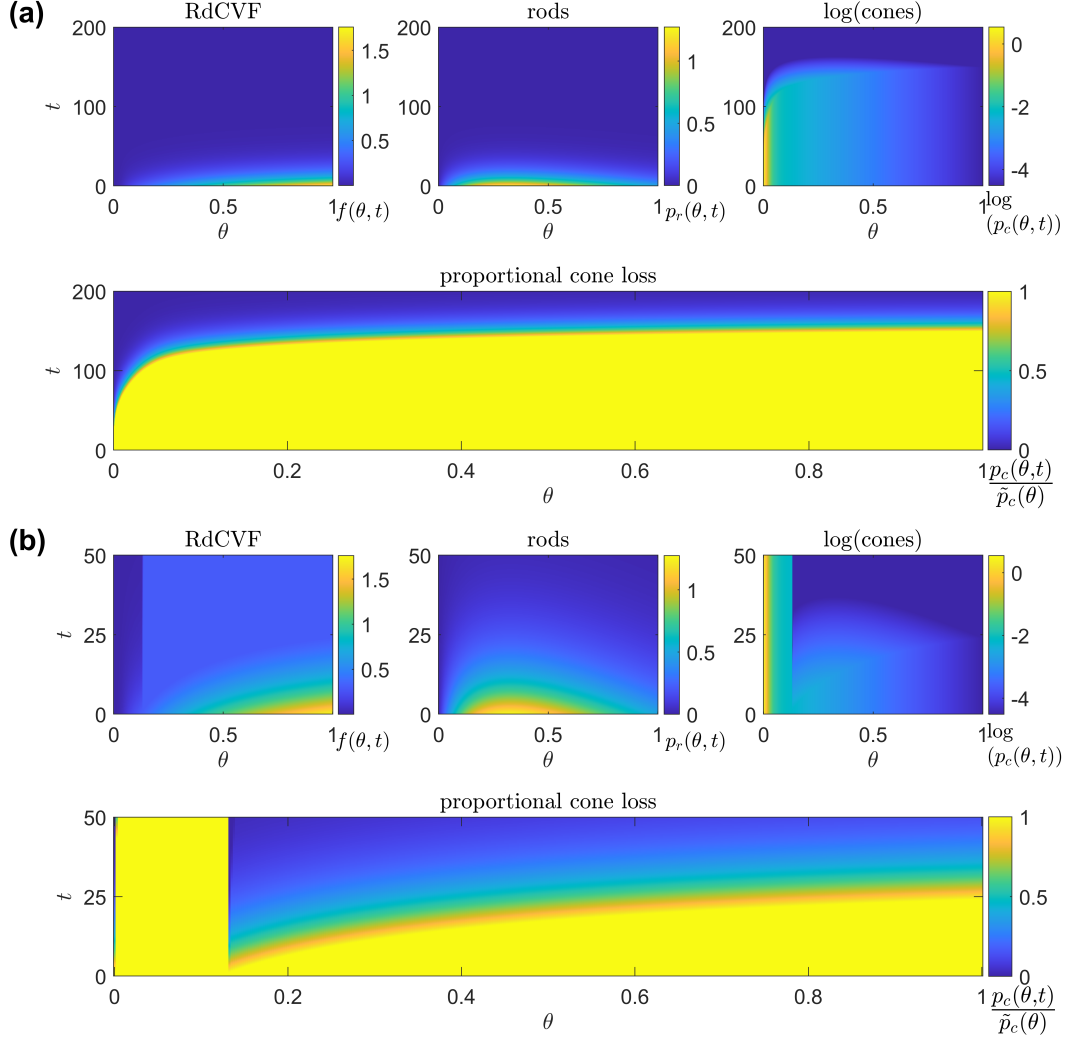


Figure 9: Simulations of RdCVF and photoreceptor dynamics with mutation-induced rod degeneration — dynamic model (Dyn Case 2). (a) and (b) top, left-to-right: TF concentration, $f(\theta, t)$, rod density, $p_r(\theta, t)$, and the natural logarithm of the cone density, $\log(p_c(\theta, t))$ (colour scale lower threshold: $\log(\min(p_{c_{\text{init}}}(\theta)))$); bottom: proportional cone loss, $p_c(\theta, t)/\bar{p}_c(\theta)$. (a) Dyn Case 2(i): $f_{\text{crit}} = 3 \times 10^{-5}$, simulation spans ~ 22.1 years in dimensional variables; (b) Dyn Case 2(ii): $f_{\text{crit}} = 0.3$, simulation spans ~ 5.5 years in dimensional variables. Cone degeneration initiates at the fovea ($\theta = 0$) in (a) and at $\theta = 0.13$ in (b), spreading peripherally (rightwards) in both cases. Eqs. (7)–(9) and (11)–(12) were solved using the method of lines, with $F(\theta) = 1$ and $G(\theta) = 1$, and without treatment or cone regeneration. (a) 4001 mesh points, (b) 401 mesh points. The parameter $\xi = 0$. Remaining parameter values as in Table 2.

Fig. 10 shows RdCVF and photoreceptor dynamics with mutation-induced rod degeneration, following the complete removal of rods from the interval $(\theta_{r_1}, \theta_{r_2})$ (Dyn Case 3). Cone degeneration initiates at the fovea ($\theta = 0$) in (a) ($f_{\text{crit}} = 3 \times 10^{-5}$) and at $\theta = 0.13$ in (b) ($f_{\text{crit}} = 0.3$), spreading peripherally (rightwards) in both cases as in Fig. 9. Mutation-induced rod loss drives cone degenerate patch expansion and precipitates the formation of cone degenerate patches where they did not previously form ((b) centre and right narrow patches).

4.2. Treatment

Fig. S4 shows the effects of treatment upon RdCVF and cone OS dynamics following the complete removal of rods from the interval $(\theta_{r_1}, \theta_{r_2})$ (Dyn Case 4). The treatment rate, ξ , is chosen to lie above the critical value, ξ_{crit} , in each case. Treatment results in the complete recovery of cone OSs in all cases, in agreement with our steady-state analytical and numerical predictions (see Figs. 5 and 6).

Fig. 11 shows the effects of treatment upon cone OS dynamics with mutation-induced rod degeneration (Dyn Case 5). Strong treatment ($\xi = 6 \times 10^4$) results in the complete recovery of cone OSs in both (a) ($f_{\text{crit}} = 3 \times 10^{-5}$) and (b) ($f_{\text{crit}} = 0.3$), while weak treatment ($\xi = 4$ in (a) and $\xi = 4 \times 10^4$ in (b)) delays cone OS degeneration in (a), and provides permanent cone OS recovery for $\theta \leq 0.13$ and temporary cone OS recovery for $\theta > 0.13$ in (b).

5. Discussion

Despite decades of research, the mechanisms underlying secondary cone loss and the spread of photoreceptor degeneration in retinitis pigmentosa (RP) have yet to be conclusively determined; though a number of candidate mechanisms have been hypothesised. One such hypothesis — the trophic factor hypothesis — suggests that rod photoreceptors produce a chemical which cone photoreceptors require to survive, such that when rods are lost, the chemical is depleted, and cone degeneration follows. Such a trophic factor (TF), rod-derived cone viability factor (RdCVF), has been chemically identified by Léveillard et al. (2004), and was found to slow the degeneration and preserve the function of cones in rat, mouse and chick models (Fintz et al., 2003; Léveillard et al., 2004; Mohand-Saïd et al., 1998, 2000, 1997; Yang et al., 2009).

In this paper, we have formulated and analysed reaction-diffusion partial differential equation (PDE) models of human RP, in which we considered RdCVF starvation as a potential mechanism of cone loss. As such, this paper complements empirical and clinical studies by isolating a biochemical mechanism in a way that would be almost impossible to achieve experimentally, allowing us to predict the *in vivo* dynamics of photoreceptor loss that would result if this mechanism (RdCVF starvation) alone were responsible for retinal degeneration (subsequent to the direct effects of the underlying mutation). We used our models to answer three key questions: (i) under what conditions will the loss of a patch of rods lead to cone degeneration?, (ii) what is the critical TF treatment dose required to prevent cone degeneration following the loss of a patch of rods?, and (iii) what spatio-temporal patterns of cone degeneration can be explained via the trophic factor mechanism alone?

We began by considering the steady-state model (Section 3), first without (Section 3.2.1) and then with (Section 3.2.2) RdCVF treatment, using a combination of numerical solutions and asymptotic analysis, achieving excellent agreement between the two.

In the untreated scenario, we considered four cases: (1) St-st Case 1 — wide patch of rod loss, (2) St-st Case 2 — narrow patch of rod loss, (3) St-st Case 3 — wide patch of rod and cone loss, and (4) St-st Case 4 — narrow patch of rod and cone loss; and for each of these cases, the two sub-cases: (i) low TF threshold concentration, $f_{\text{crit}} = 3 \times 10^{-5}$, and (ii) high TF threshold concentration, $f_{\text{crit}} = 0.3$ (Figs. S2, 3 and 4). In all cases it was found that cone degenerate patch width increases monotonically with increasing rod degenerate

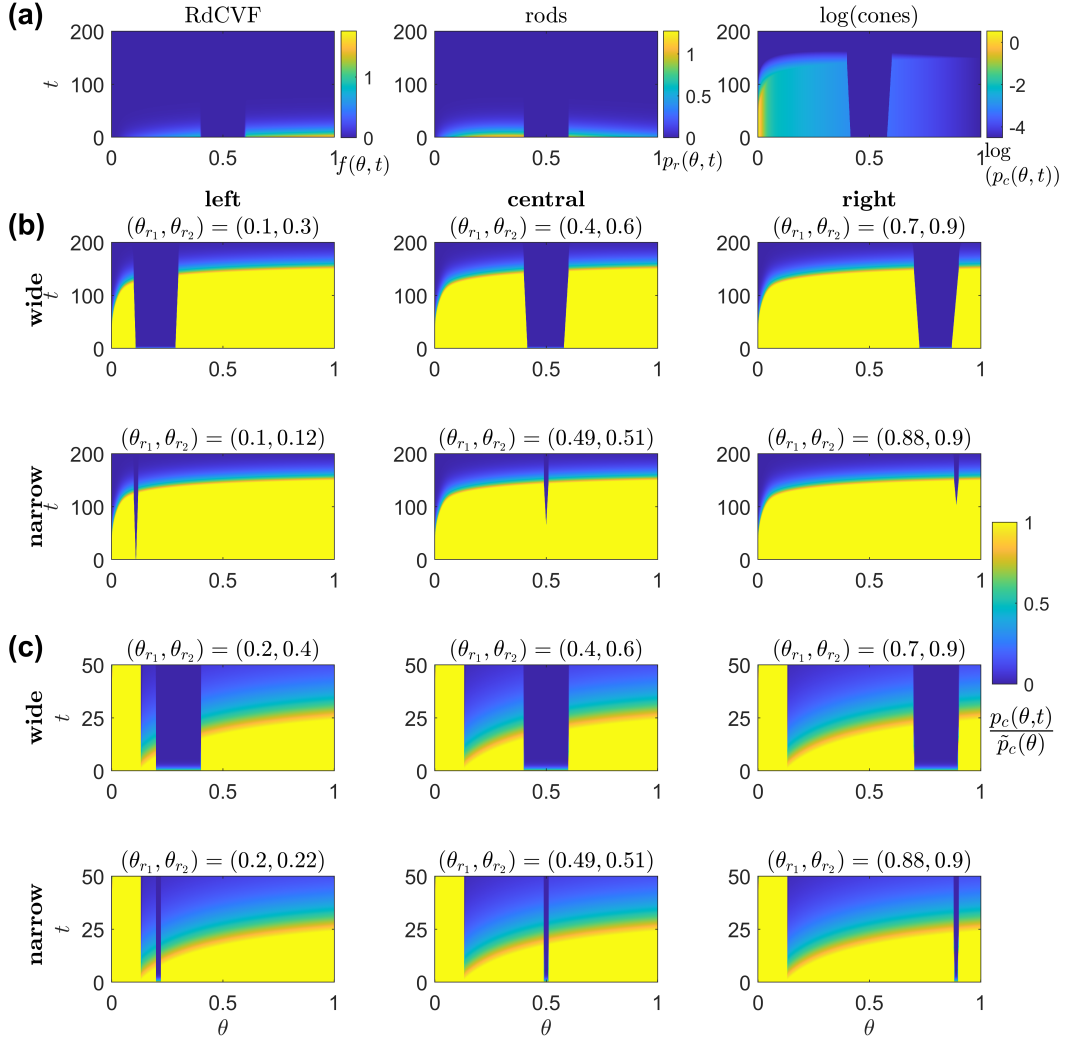


Figure 10: Simulations of RdCVF and photoreceptor dynamics with mutation-induced rod degeneration, following the complete removal of rods from the interval $(\theta_{r_1}, \theta_{r_2})$ — dynamic model (Dyn Case 3). (a) left-to-right: TF concentration, $f(\theta, t)$, rod density, $p_r(\theta, t)$, and the natural logarithm of the cone density, $\log(p_c(\theta, t))$ (colour scale lower threshold: $\log(\min(p_{c_{\text{init}}}(\theta)))$), for Dyn Case 3(i): $f_{\text{crit}} = 3 \times 10^{-5}$ and $(\theta_{r_1}, \theta_{r_2}) = (0.4, 0.6)$ (the same as for the top-middle panel in (b)). (b) proportional cone loss, $p_c(\theta, t)/\bar{p}_c(\theta)$, for Dyn Case 3(i): $f_{\text{crit}} = 3 \times 10^{-5}$. (c) proportional cone loss for Dyn Case 3(ii): $f_{\text{crit}} = 0.3$. Rods are removed from wide (width $O(1)$) and narrow (width $O(\epsilon)$) intervals, on the left-hand side (parafovea/perifovea), centre (mid-periphery) and right-hand side (far-periphery) of the domain. (a) and (b) simulation spans ~ 22.1 years in dimensional variables, (c) simulation spans ~ 5.5 years in dimensional variables. Cone degeneration initiates at the fovea ($\theta = 0$) in (a) and (b), and at $\theta = 0.13$ in (c), spreading peripherally (rightwards) in both cases. Patches of cone loss form and expand in all cases, initiating within patches of rod loss (compare with Fig. 8). Eqs. (7)–(9) and (11)–(12) were solved using the method of lines, with $F(\theta) = H(\theta_{r_1} - \theta) + H(\theta - \theta_{r_2})$ and $G(\theta) = 1$, and without treatment or cone regeneration. (a) and (b) 4001 mesh points, (c) 401 mesh points. The parameter $\xi = 0$. Remaining parameter values as in Table 2.

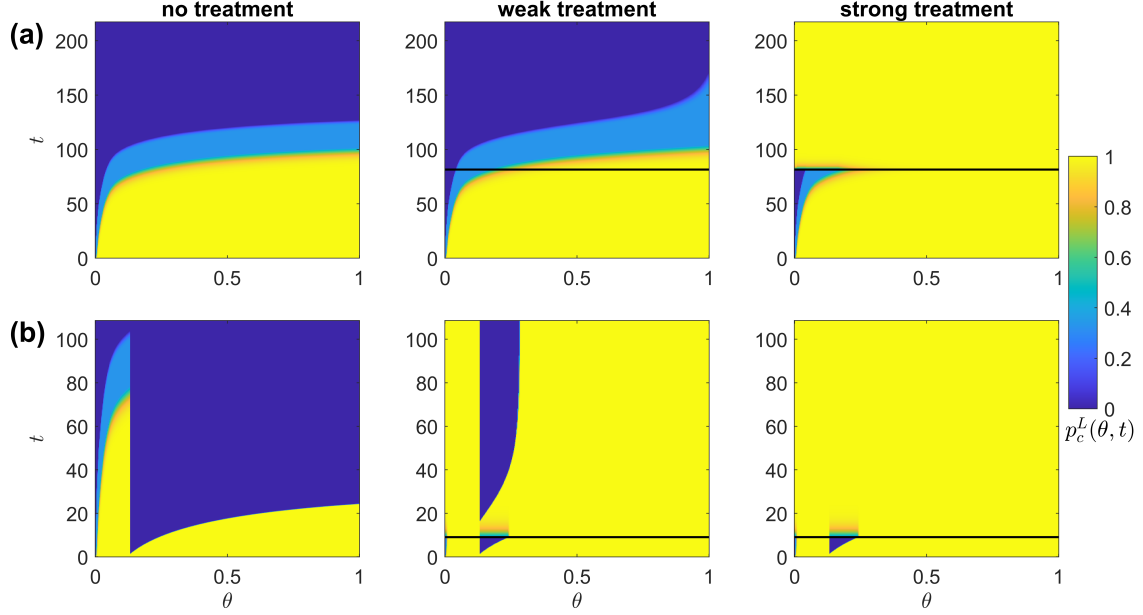


Figure 11: Simulations of the effects of treatment upon cone OS dynamics with mutation-induced rod degeneration — dynamic model (Dyn Case 5). Panels show cone OS length, $p_c^L(\theta, t)$, with no treatment (column 1), weak treatment (column 2) and strong treatment (column 3). Black horizontal lines mark the time point, t_{crit} , at which treatment is introduced. (a) Dyn Case 5(i): $f_{\text{crit}} = 3 \times 10^{-5}$ and $t_{\text{crit}} = 81.5$, while the simulation spans a period of 24 years in dimensional variables; (b) Dyn Case 5(ii): $f_{\text{crit}} = 0.3$ and $t_{\text{crit}} = 9.05$, while the simulation spans a period of 12 years in dimensional variables. Strong treatment results in the complete recovery of cone OSs in both (a) and (b), while weak treatment delays cone OS degeneration in (a), and provides permanent cone OS recovery for $\theta \leq 0.13$ and temporary cone OS recovery for $\theta > 0.13$ in (b). Eqs. (7), (8) and (10)–(12) were solved using the method of lines, with 401 mesh points, $F(\theta) = 1$, $G(\theta) = 1$ and $T(\theta, t) = H(t - t_{\text{crit}})$. Parameter values: (a) and (b) column 1, $\xi = 0$; (a) column 2, $\xi = 4$; (b) column 2, $\xi = 4 \times 10^4$; (a) and (b) column 3, $\xi = 6 \times 10^4$. Remaining parameter values as in Table 2.

patch width and with decreasing rod degenerate patch centre eccentricity, while the difference between rod and cone degenerate patch widths decreases monotonically with rod degenerate patch centre eccentricity. Cone degenerate patch width was predicted to be less than rod degenerate patch width, except for wide and narrow rod loss patches with $f_{\text{crit}} = 0.3$ (St-st Cases 1(ii) and 2(ii)), where the cone degenerate patch width may exceed rod degenerate patch width close to the fovea. These results make sense in the light of the fact that the ratio of rods to cones increases monotonically with increasing eccentricity (Fig. 2(a)), such that TF is scarcer toward the centre of the retina, leading to the formation of larger cone degenerate patches. It would be interesting to test these predictions experimentally by measuring the cone density local to patches of rod loss.

In the treated scenario, we considered four cases: (1) St-st Case 5 — wide patch of rod loss with global treatment, (2) St-st Case 6 — wide patch of rod loss with local treatment, (3) St-st Case 7 — narrow patch of rod loss with global treatment, and (4) St-st Case 8 — narrow patch of rod loss with local treatment; and for each of these cases, the two sub-cases: (i) $f_{\text{crit}} = 3 \times 10^{-5}$, and (ii) $f_{\text{crit}} = 0.3$ (Figs. 5–7 and S3). In all cases the critical treatment rate, ξ_{crit} , increases monotonically with decreasing rod degenerate patch centre and left boundary eccentricity. As above, this is to be expected from consideration of the ratio of rods to cones (Fig. 2(a)), as endogenously produced TF is scarcer towards the retinal centre.

In the wide patch cases (St-st Cases 5 and 6), the critical treatment rate depends almost entirely upon the rod degenerate patch left boundary position, while being independent of rod degenerate patch width (Fig. 5). By contrast, in the narrow patch cases (St-st Cases 7 and 8), the critical treatment rate depends upon both the rod degenerate patch left and right boundary positions, and hence upon the rod degenerate patch width, the critical treatment rate increasing monotonically with increasing rod degenerate patch width (Fig. 6). The reason for this difference becomes clear from considering the normalised eccentricity of the minimum TF concentration, $\hat{\theta}_{\text{crit}}$, in Fig. S3. For wide patches (Fig. S3(a)–(d)), $\hat{\theta}_{\text{crit}}$ is close to the left-hand rod degenerate patch boundary, θ_{r_1} , such that $\hat{\theta}_{\text{crit}} \in (0, 0.2)$ is in the left-centre-inner region (Fig. A.1(a)(ii)), where only the left-hand boundary exerts an influence. Therefore, ξ_{crit} only depends on the value of θ_{r_1} in this case. Whereas, for narrow patches (Fig. S3(e)–(f)), $\hat{\theta}_{\text{crit}}$ is close to the centre of the rod degenerate patch, such that $\hat{\theta}_{\text{crit}} \sim 0.5$ is in the centre-inner region (Fig. A.1(b)(ii)), upon which both boundaries exert an influence. Therefore, ξ_{crit} depends on the values of both θ_{r_1} and θ_{r_2} in this case.

The normalised eccentricity of the minimum TF concentration, $\hat{\theta}_{\text{crit}}$, is close to the left- rather than the right-hand rod degenerate patch boundary in the wide patch case because the rod to cone ratio, and hence the TF concentration, is lower there (Fig. 2(a)). $\hat{\theta}_{\text{crit}}$ increases monotonically with decreasing rod degenerate patch width and with increasing rod degenerate patch centre and left boundary position in all cases. Knowing the location of $\hat{\theta}_{\text{crit}}$ is important as it tells us which region of the retina is most at risk of degeneration (i.e. which area will degenerate first), suggesting where to spatially target treatment (e.g. with sub-retinal RdCVF injections).

Trophic factor (RdCVF) treatment could be administered in a number of ways. RdCVF could be injected either into the vitreous (the structure filling the centre of the eye), or into the sub-retinal space between the photoreceptors and the retinal pigment epithelium (RPE; Yang et al., 2009). Injection into the vitreous is less invasive and would distribute RdCVF across the whole retina, while sub-retinal injections would target specific retinal areas and allow for a much higher dose to reach the cones (Yang et al., 2009). A downside to injections is that they would have to be regularly repeated throughout the lifetime of a patient, to maintain an adequate supply of RdCVF (Clérin et al., 2020). An alternative mode of administration might be to use encapsulated cell-based delivery, in which a polymer membrane capsule loaded with cells genetically modified to secrete TF is surgically implanted into the vitreous, releasing TF over a period of months to years (Musarella and MacDonald, 2011; Sieving et al., 2006; Tao et al., 2002). This would require less frequent interventions than with injections and provide a constant supply of RdCVF. This procedure has

been tested in canines and humans, using ciliary neurotrophic factor producing RPE cells, and was found to slow photoreceptor degeneration and preserve visual acuity (Sieving et al., 2006; Tao et al., 2002). Lastly, retinal gene therapy, administered through intravitreal or sub-retinal injection of appropriately engineered adeno-associated virus vectors, could be used to genetically modify retinal cells other than the degenerating rods (e.g. RPE cells) to produce RdCVF, providing a constant, endogenous supply for the cones (Byrne et al., 2015; Clérin et al., 2020).

Comparison of critical treatment rates in the global and local treatment cases (St-st Cases 5–8) shows that ξ_{crit} is almost identical for global and local treatment in the wide patch cases (St-st Cases 5 and 6), while the critical treatment rate is higher for local than for global treatment for the narrowest patches close to the fovea in the narrow patch cases (St-st Cases 7 and 8; Fig. 7). Therefore, a targeted treatment (e.g. a localised sub-retinal injection of RdCVF), which uses less TF, could be just as effective as a global treatment in most cases. It would be interesting to test these and the above treatment-related predictions experimentally, comparing the effects of different RdCVF doses, injection locations and local vs. global treatments.

Next we considered numerical solutions to the dynamic, time-dependent model (Section 4), first without (Section 4.1) and then with (Section 4.2) RdCVF treatment.

In the untreated scenario we considered three cases: (1) Dyn Case 1 — wide and narrow patches of rod loss without mutation-induced rod degeneration, (2) Dyn Case 2 — mutation-induced rod degeneration without patch loss, and (3) Dyn Case 3 — wide and narrow patches of rod loss with mutation-induced rod degeneration; and for each of these cases, the two sub-cases: (i) $f_{\text{crit}} = 3 \times 10^{-5}$, and (ii) $f_{\text{crit}} = 0.3$ (Figs. 8–10). In Dyn Case 1, it was found that cone degeneration always occurs within wide rod degenerate patches, whereas cone degeneration may not occur in narrow rod degenerate patches away from the retinal centre for $f_{\text{crit}} = 3 \times 10^{-5}$ (Fig. 8). Further, for $f_{\text{crit}} = 0.3$, the spatial extent of cone loss may exceed that of rod loss in near-central rod degenerate patches. This is in good agreement with our predictions from the steady-state problem in Section 3 (Figs. 3 and 4).

In Dyn Case 2 we consider the question ‘what spatio-temporal patterns of cone degeneration can be explained via the TF mechanism alone?’ It was found that cone degeneration initiates at the retinal centre and spreads peripherally, accelerating as it does so (Fig. 9). This does not replicate any of the spatio-temporal patterns of visual field loss (and hence retinal degeneration) described by Grover et al. (1998) (see also Roberts et al., 2018). In particular, it fails to replicate the sparing of the central retina that is usually observed. This suggests that other mechanisms are necessary, either in place of or (more likely) together with trophic factor starvation-induced degeneration, to explain *in vivo* disease progression. However, we note that other spatio-temporal patterns could be generated using our model by making alternative parameter choices. For example, the rates of TF production, α , and consumption, β , could be reduced by two orders of magnitude, bringing the TF decay term into the dominant balance with the production and consumption terms in Eqn. (7). Further, it may be that the TF threshold concentration, f_{crit} , and the rate of mutation-induced rod degeneration, ϕ_r , vary with eccentricity. For example, f_{crit} may be lower at the retinal centre if the cones in that region are protected in some way from the low RdCVF levels found there, while RdCVF supply to the central retina would be maintained for longer if ϕ_r were reduced in that region (ϕ_r was shown to vary with eccentricity in the healthy ageing retina by Curcio et al., 1993, thus it is reasonable to assume that it would also vary in the diseased retina, though we note that in the healthy case it is the central rods that are most vulnerable). We will explore the effects of these alternative parameter choices in a future publication.

In Dyn Case 3, mutation-induced rod degeneration was shown to precipitate the formation of cone degenerate patches at the locations of narrow rod degenerate patches, where they would not have otherwise formed, and to drive the expansion of cone degenerate patches (Fig. 10). This occurs because the loss of rods reduces the rate of production of TF, pushing RdCVF concentrations below the critical threshold in regions which were previously well-supplied.

As a hypothesis to explain the spread of photoreceptor degeneration, the trophic factor mechanism is *self-limiting* when considered in isolation and in the absence of mutation-induced rod degeneration. This is because, following the loss of a patch of rods (and consequent drop in local RdCVF concentration) in an otherwise healthy retina, the loss of cones decreases TF demand until RdCVF levels rise above the threshold concentration. This is in contrast to the oxygen toxicity mechanism, considered by the author in previous publications (see Roberts et al., 2017, 2018), which is *self-reinforcing*; toxically high oxygen levels causing photoreceptor degeneration, leading to a further rise in oxygen levels, and so on.

In the treated scenario we considered two cases: (1) Dyn Case 4 — wide and narrow patches of rod loss, without mutation-induced rod degeneration, with global and local treatment, and (2) Dyn Case 5 — mutation-induced rod degeneration, without patch loss, with weak and strong global treatment; and for each of these cases, the two sub-cases: (i) $f_{\text{crit}} = 3 \times 10^{-5}$, and (ii) $f_{\text{crit}} = 0.3$ (Figs. S4 and 11). In Dyn Case 4, the treatment rate, ξ , was chosen to lie above the critical treatment rate, ξ_{crit} , predicted from the steady-state model (Figs. 5 and 6), resulting in the complete recovery of cone outer segments (OSs) in all cases as expected (Fig. S4). In Dyn Case 5, strong treatment ($\xi = 6 \times 10^4$) resulted in the complete recovery of cone OSs, while weak treatment ($\xi = 4$ or $\xi = 4 \times 10^4$) delayed or temporarily reversed cone OS degeneration (Fig. 11). While the possible rates of RdCVF treatment have yet to be measured, our models provide useful, experimentally testable predictions of the critical treatment rate required to prevent or reverse damage to cones and the possible effects of different treatment rates.

Throughout this paper we have focussed on the human retina, incorporating its distinctive photoreceptor distribution, so as to ensure clinical relevance. However, as noted in Section 1, model animal RP species, such as mice, rats and pigs, do not have foveated retinas, their rod and cone densities varying more gradually across the eye (Chandler et al., 1999; Gaillard et al., 2009; Ortín-Martínez et al., 2014). Since the healthy rod and cone densities determine the model behaviour, it would be of great interest to explore the effect of these alternative photoreceptor distributions in future studies.

In future work, we will extend the present study in four main directions. First, we will explore alternative scalings on the rates of RdCVF production, α , and consumption, β , together with the effects of spatially heterogeneous TF threshold concentration, f_{crit} , and rate of mutation-induced rod degeneration, ϕ_r , upon spatio-temporal patterns of retinal degeneration, as discussed above. Second, we will extend our models to two spatial dimensions, covering the region between the fovea/(retinal centre) and the ora serrata and incorporating variations in the azimuthal dimension (as in Roberts et al., 2018). Third, we will expand our study to include animal retinas, accounting for the effects of different photoreceptor distributions upon disease dynamics (e.g. mouse retinas, which exhibit the same initial patchy loss of photoreceptors as seen in humans; García-Ayuso et al., 2013; Ji et al., 2012; Lee et al., 2011; Zhu et al., 2013). Fourth, we will incorporate further biochemical details, such as those considered in Camacho et al. (2019).

In conclusion, we have constructed novel mathematical models of RP to describe and predict retinal degeneration, and the effects of treatment, under the trophic factor hypothesis. We predicted the spatial extent of cone degeneration following the loss of a patch of rods, and the critical RdCVF treatment rate required to prevent cone loss in this case. Lastly, we predicted the spatio-temporal patterns of cone degeneration that would result from this mechanism in isolation, suggesting that additional mechanisms are required to explain the patterns seen *in vivo*.

Appendix A. Asymptotic analysis — St-st Case 1: wide patch of rod loss without treatment

In this section we perform an asymptotic analysis with the aim of deriving analytical expressions for the boundaries of cone loss following the formation of a degenerate rod patch. We assume that cone loss will occur in the region for which the TF concentration $f < f_{\text{crit}}$ local to the degenerate rod patch. Thus, the

boundaries of the degenerate cone patch will be given as the eccentricities at which $f = f_{\text{crit}}$. To determine where this occurs, we seek a leading order solution for $f(\theta)$ to Eqs. (7) and (11) at steady-state, for prescribed $p_r(\theta) = F(\theta)\tilde{p}_r(\theta)$ and $p_c(\theta) = G(\theta)\tilde{p}_c(\theta)$, where $F(\theta) = H(\theta_{r_1} - \theta) + H(\theta - \theta_{r_2})$ and $G(\theta) = 1$.

In what follows, we use the following variables and parameters, defined here for ease of reference:

- $\tilde{p}_{c_0}(\theta) = B_2 e^{-b_2 \theta}$;
- $\tilde{p}_{r_0}(\theta) = B_3 \theta e^{-b_3 \theta}$;
- $c_1 := \beta \tilde{p}_{c_0}(\theta_{r_1}) / D_f = \beta B_2 e^{-b_2 \theta_{r_1}} / D_f$;
- $\bar{c}_1 := \beta \tilde{p}_{c_0}(\theta_{r_2}) / D_f = \beta B_2 e^{-b_2 \theta_{r_2}} / D_f$;
- $c_2 := \alpha \tilde{p}_{r_0}(\theta_{r_1}) / D_f = \alpha B_3 \theta_{r_1} e^{-b_3 \theta_{r_1}} / D_f$;
- $\bar{c}_2 := \alpha \tilde{p}_{r_0}(\theta_{r_2}) / D_f = \alpha B_3 \theta_{r_2} e^{-b_3 \theta_{r_2}} / D_f$.

We choose $\epsilon = O(10^{-2})$, noting that we set $\epsilon = 10^{-2}$ for all plots, scaling parameters as follows: $\alpha = \epsilon^{-2} \alpha'$, $\beta = \epsilon^{-3} \beta'$, $\eta = \epsilon^{-1} \eta'$ and $b_1 = \epsilon^{-1} b_1'$ (using the standard parameter values as in Table 2). All remaining parameters are assumed to be $O(1)$. We note that we could have chosen values for α and β two orders of magnitude smaller than their present values, resulting in an alternative distinguished limit. This alternative scaling will be explored in a future publication. We also scale the dependent variable $p_c = \epsilon p'_c$, and assume $f = O(1)$ and $p_r = O(1)$. This scaling on the dependent variables is valid in the region $\theta \in (\epsilon, 1]$ here and in all rod loss only cases. For completeness, we note that there are two further regions with different scalings on the dependent variables in the interval $\theta \in [0, \epsilon)$. Analysis within these regions is beyond the scope of this study.

Applying the above scalings to the steady-state version of Eqn. (7) and dropping the dashes, we obtain

$$0 = \epsilon^2 D_f \frac{d^2 f}{d\theta^2} + \epsilon^2 D_f \Theta \cot(\Theta \theta) \frac{df}{d\theta} + \alpha p_r - \beta p_c f - \epsilon \eta f, \quad (\text{A.1})$$

where,

$$\tilde{p}_r(\theta) = B_3 \theta e^{-b_3 \theta}, \quad (\text{A.2})$$

$$\tilde{p}_c(\theta) = B_1 e^{-\epsilon^{-1} b_1 \theta} + B_2 e^{-b_2 \theta}. \quad (\text{A.3})$$

We note that $\cot(\Theta \theta) \leq O(1)$ for $\theta \in (0.1, 1]$ and $\theta_{r_2} - \theta_{r_1} \leq O(1)$. Therefore, the following analysis is valid in the region $\theta \in (0.1, 1 - \epsilon)$ (where we subtract ϵ from the right-hand boundary to avoid the boundary layer there).

We close the system by imposing zero-flux boundary conditions as in Eqn. (11):

$$\frac{df}{d\theta}(0) = 0 = \frac{df}{d\theta}(1). \quad (\text{A.4})$$

Next, we decompose the domain into left ($\theta < \theta_{r_1}$), centre ($\theta_{r_1} < \theta < \theta_{r_2}$) and right ($\theta > \theta_{r_2}$), outer (width $O(1)$) and inner (width $O(\epsilon)$) regions as depicted in Fig. A.1(a). Discontinuities are introduced into Eqn. (A.1) at the edges of the degenerate rod patch (θ_{r_1} and θ_{r_2}), accounting for the left/centre/right division, while boundary layers are present either side of these discontinuities, where there is a sharp transition in $f(\theta)$, and also at the ends of the domain, to satisfy the zero-flux boundary conditions. In the following analysis we are not interested in the behaviour of the solution within the boundary layers at the ends of the

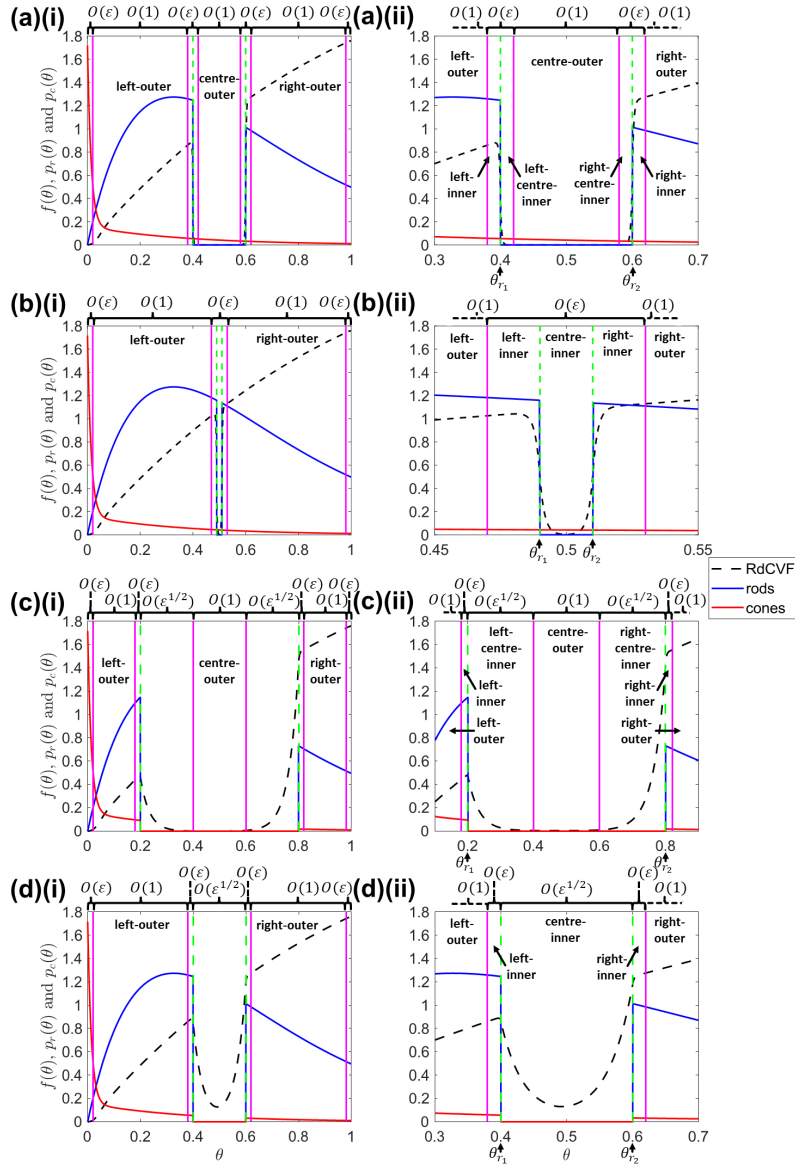


Figure A.1: Inner and outer region locations for the asymptotic analysis. (a) St-st Cases 1, 5 and 6: wide (width $O(1)$) patch of rod loss (untreated RdCVF profile plotted here); (b) St-st Cases 2, 7 and 8: narrow (width $O(\epsilon)$) patch of rod loss (untreated RdCVF profile plotted here); (c) St-st Case 3: wide (width $O(1)$) patch of rod and cone loss; (d) St-st Case 4: narrow (width $O(\epsilon^{1/2})$) patch of rod and cone loss. The full domain is shown in the left-hand column, while the right-hand column magnifies the region from which photoreceptors have been removed. The green dashed vertical lines mark the boundaries of rod (and cone) loss, while the solid magenta vertical lines demarcate the approximate limits of the boundary layers. To obtain $f(\theta)$, Eqs. (7) and (11) were solved using the finite difference method, with 4001 mesh points, where $p_r(\theta) = F(\theta)\bar{p}_r(\theta)$, $p_c(\theta) = G(\theta)\bar{p}_c(\theta)$, $F(\theta) = H(\theta_{r_1} - \theta) + H(\theta - \theta_{r_2})$, $G(\theta) = 1$ in (a) and (b), and $G(\theta) = H(\theta_{c_1} - \theta) + H(\theta - \theta_{c_2})$ in (c) and (d), and without treatment, cone regeneration or mutation-induced rod degeneration. Parameter values: $\xi = 0$; (a) $(\theta_{r_1}, \theta_{r_2}) = (0.4, 0.6)$; (b) $(\theta_{r_1}, \theta_{r_2}) = (0.49, 0.51)$; (c) $(\theta_{r_1}, \theta_{r_2}) = (\theta_{c_1}, \theta_{c_2}) = (0.2, 0.8)$; and (d) $(\theta_{r_1}, \theta_{r_2}) = (\theta_{c_1}, \theta_{c_2}) = (0.4, 0.6)$. Remaining parameter values as in Table 2.

domain. We label the remaining regions as follows from left to right: left-outer, left-inner, left-centre-inner, centre-outer, right-centre-inner, right-inner and right-outer (see Fig. A.1(a)(ii)).

In what follows we seek leading order ($O(1)$) solutions to $f(\theta)$ in each of the outer and inner regions. This results in a number of unknown constants of integration, whose values are determined by a combination of matching and patching. Asymptotic matching ensures that the inner limit of the outer solutions match with the outer limit of the inner solutions where they meet, while patching ensures continuity of TF concentration and flux across the discontinuities at θ_{r_1} and θ_{r_2} (Bender and Orszag, 1999).

We form the regular perturbation expansions:

$$f(\theta) = f_0(\theta) + \epsilon f_1(\theta) + O(\epsilon^2), \quad (\text{A.5})$$

$$p_r(\theta) = p_{r_0}(\theta) + \epsilon p_{r_1}(\theta) + O(\epsilon^2), \quad (\text{A.6})$$

$$p_c(\theta) = p_{c_0}(\theta) + \epsilon p_{c_1}(\theta) + O(\epsilon^2), \quad (\text{A.7})$$

and seek $f_0(\theta)$, where $p_{r_0}(\theta) = F(\theta)\tilde{p}_{r_0}(\theta) = (H(\theta_{r_1} - \theta) + H(\theta - \theta_{r_2}))B_3\theta e^{-b_3\theta}$ and $p_{c_0}(\theta) = G(\theta)\tilde{p}_{c_0}(\theta) = B_2e^{-b_2\theta}$.

Seeking leading order solutions to (A.1) in the left- and right-outer regions, we obtain,

$$f_0(\theta) = \frac{\alpha B_3 \theta e^{-b_3 \theta}}{\beta B_2 e^{-b_2 \theta}},$$

while, in the centre-outer region, we obtain,

$$f_0(\theta) = 0.$$

To find the leading order solution in the left-inner and left-centre-inner regions, we rescale the independent variable θ as $\hat{\theta} = \epsilon^{-1}(\theta - \theta_{r_1})$, such that the first diffusion term enters the dominant balance. Thus, Eqn. (A.1) becomes

$$0 = D_f \frac{d^2 f}{d\hat{\theta}^2} + \epsilon D_f \Theta \cot(\Theta(\theta_{r_1} + \epsilon\hat{\theta})) \frac{df}{d\hat{\theta}} + \alpha p_r - \beta p_c f - \epsilon \eta f. \quad (\text{A.8})$$

Thus, we obtain leading order solutions in the left-inner region,

$$f(\hat{\theta}) = A_1 e^{\sqrt{c_1}\hat{\theta}} + A_2 e^{-\sqrt{c_1}\hat{\theta}} + \frac{c_2}{c_1}, \quad (\text{A.9})$$

and in the left-centre-inner region,

$$f(\hat{\theta}) = A_3 e^{\sqrt{c_1}\hat{\theta}} + A_4 e^{-\sqrt{c_1}\hat{\theta}}, \quad (\text{A.10})$$

where $c_1 = \beta \tilde{p}_{c_0}(\theta_{r_1})/D_f = \beta B_2 e^{-b_2 \theta_{r_1}}/D_f$, $c_2 = \alpha \tilde{p}_{r_0}(\theta_{r_1})/D_f = \alpha B_3 \theta_{r_1} e^{-b_3 \theta_{r_1}}/D_f$, and A_1 – A_4 are constants of integration to be determined.

To find the leading order solution in the right-centre-inner and right-inner regions, we rescale the independent variable θ as $\bar{\theta} = \epsilon^{-1}(\theta - \theta_{r_2})$, obtaining the equivalent equation to Eqn. (A.8), but with $\hat{\theta} \mapsto \bar{\theta}$ and $\theta_{r_1} \mapsto \theta_{r_2}$.

This gives us leading order solutions in the right-centre-inner region,

$$f(\bar{\theta}) = A_5 e^{\sqrt{c_1}\bar{\theta}} + A_6 e^{-\sqrt{c_1}\bar{\theta}}, \quad (\text{A.11})$$

and in the right-inner region,

$$f(\bar{\theta}) = A_7 e^{\sqrt{c_1}\bar{\theta}} + A_8 e^{-\sqrt{c_1}\bar{\theta}} + \frac{\bar{c}_2}{\bar{c}_1}, \quad (\text{A.12})$$

where $\bar{c}_1 = \beta \tilde{p}_{c_0}(\theta_{r_2})/D_f = \beta B_2 e^{-b_2 \theta_{r_2}}/D_f$, $\bar{c}_2 = \alpha \tilde{p}_{r_0}(\theta_{r_2})/D_f = \alpha B_3 \theta_{r_2} e^{-b_3 \theta_{r_2}}/D_f$, and A_5 – A_8 are constants of integration to be determined.

To obtain values for A_1 – A_8 we first match and then patch. We match using Van Dyke's matching rule, which states that the m term inner expansion of the n term outer solution, should equal the n term outer expansion of the m term inner solution. In this case, since we are dealing with leading order solutions only, $m = n = 1$. Matching the left-outer and left-inner solutions, we form the one term inner expansion of the one term outer solution,

$$f(\hat{\theta}) \sim \frac{\alpha B_3 \theta_{r_1} e^{-b_3 \theta_{r_1}}}{\beta B_2 e^{-b_2 \theta_{r_1}}} = \frac{c_2}{c_1},$$

and the one term outer expansion of one term inner solution,

$$f(\theta) \sim A_1 e^{\sqrt{c_1} \epsilon^{-1}(\theta - \theta_{r_1})} + A_2 e^{-\sqrt{c_1} \epsilon^{-1}(\theta - \theta_{r_1})} + \frac{c_2}{c_1}.$$

Since $\theta - \theta_{r_1} > 0$ in the left-hand regions, the first term is exponentially small and so can be neglected, while the second term is exponentially large and so we must have $A_2 = 0$. Matching the left-centre-inner and centre-outer solutions, the centre-outer and right-centre-inner solutions, and the right-inner and right-outer solutions in a similar fashion, we obtain $A_3 = 0$, $A_6 = 0$, and $A_7 = 0$ respectively.

Patching across $\theta = \theta_{r_1}$ and $\theta = \theta_{r_2}$, we obtain expression for A_1 , A_4 , A_5 and A_8 in terms of known parameters. Substituting for A_1 – A_8 into Eqs. (A.9)–(A.12), we obtain the full set of leading order outer and inner solutions as follows —

Left- and Right-Outer:

$$f_0(\theta) = \frac{\alpha B_3 \theta e^{-b_3 \theta}}{\beta B_2 e^{-b_2 \theta}}, \quad (\text{A.13})$$

Left-Inner:

$$f_0(\theta) = \frac{c_2}{c_1} \left(1 - \frac{1}{2} e^{\epsilon^{-1} \sqrt{c_1}(\theta - \theta_{r_1})} \right), \quad (\text{A.14})$$

Left-Centre-Inner:

$$f_0(\theta) = \frac{c_2}{2c_1} e^{-\epsilon^{-1} \sqrt{c_1}(\theta - \theta_{r_1})}, \quad (\text{A.15})$$

Centre-Outer:

$$f_0(\theta) = 0, \quad (\text{A.16})$$

Right-Centre-Inner:

$$f_0(\theta) = \frac{\bar{c}_2}{2\bar{c}_1} e^{\epsilon^{-1} \sqrt{\bar{c}_1}(\theta - \theta_{r_2})}, \quad (\text{A.17})$$

Right-Inner:

$$f_0(\theta) = \frac{\bar{c}_2}{\bar{c}_1} \left(1 - \frac{1}{2} e^{-\epsilon^{-1} \sqrt{\bar{c}_1}(\theta - \theta_{r_2})} \right). \quad (\text{A.18})$$

Next we calculate leading order composite solutions, providing uniformly valid expansions across the left-outer and left-inner regions (left-composite); across the left-centre-inner, centre-outer and right-centre-inner regions (centre-composite), and across the right-inner and right-outer regions (right-composite). Composite solutions are calculated by adding together the relevant outer and inner solutions and subtracting one copy of the common terms to avoid double counting (see Bender and Orszag, 1999, for further details). We obtain the following composite solutions —

Left-Composite:

$$f_{0_{left-comp}}(\theta) = \frac{\alpha B_3 \theta e^{-b_3 \theta}}{\beta B_2 e^{-b_2 \theta}} - \frac{c_2}{2c_1} e^{\epsilon^{-1} \sqrt{c_1}(\theta - \theta_{r_1})}, \quad (\text{A.19})$$

Centre-Composite:

$$f_{0_{centre-comp}}(\theta) = \frac{c_2}{2c_1} e^{-\epsilon^{-1} \sqrt{c_1}(\theta - \theta_{r_1})} + \frac{\bar{c}_2}{2\bar{c}_1} e^{\epsilon^{-1} \sqrt{\bar{c}_1}(\theta - \theta_{r_2})}, \quad (\text{A.20})$$

Right-Composite:

$$f_{0_{right-comp}}(\theta) = \frac{\alpha B_3 \theta e^{-b_3 \theta}}{\beta B_2 e^{-b_2 \theta}} - \frac{\bar{c}_2}{2\bar{c}_1} e^{-\epsilon^{-1} \sqrt{\bar{c}_1}(\theta - \theta_{r_2})}. \quad (\text{A.21})$$

Lastly, we derive expressions for the cone degenerate patch boundaries. As discussed above, we assume that the boundaries of a cone degenerate patch, θ_{crit_1} (left-hand boundary) and θ_{crit_2} (right-hand boundary), occur where $f(\theta) = f_{crit}$. This cannot occur in the outer regions, since we have chosen f_{crit} such that $f(\theta) > f_{crit}$ in the healthy retina (left- and right-outer regions), while $f_0(\theta) = 0 < f_{crit}$ in the centre-outer region. Therefore, we must have $f(\theta) = f_{crit}$ somewhere in the left-inner or left-centre-inner region (θ_{crit_1}), and somewhere in the right-centre-inner or right-inner region (θ_{crit_2}). Setting $f(\theta) = f_{crit}$ in Eqs. (A.14), (A.15), (A.17) and (A.18), and rearranging, we obtain —

Minimal Cone Degenerate Patch Left Boundary:

$$\theta_{crit_1} = \theta_{r_1} + \text{sgn}\left(f_{crit} - \frac{c_2}{2c_1}\right) \frac{\epsilon}{\sqrt{c_1}} \log\left(1 - \left|\frac{2c_1 f_{crit}}{c_2} - 1\right|\right), \quad (\text{A.22})$$

Maximal Cone Degenerate Patch Right Boundary:

$$\theta_{crit_2} = \theta_{r_2} - \text{sgn}\left(f_{crit} - \frac{\bar{c}_2}{2\bar{c}_1}\right) \frac{\epsilon}{\sqrt{\bar{c}_1}} \log\left(1 - \left|\frac{2\bar{c}_1 f_{crit}}{\bar{c}_2} - 1\right|\right), \quad (\text{A.23})$$

where $\text{sgn}(\cdot)$ is the signum (or sign) function, defined such that,

$$\text{sgn}(x) := \begin{cases} -1 & \text{if } x < 0, \\ 0 & \text{if } x = 0, \\ 1 & \text{if } x > 0. \end{cases}$$

We have combined the expressions for θ_{crit_1} resulting from Eqs. (A.14) and (A.15), to give Eqn. (A.22), and similarly for θ_{crit_2} with Eqs. (A.17) and (A.18) to give Eqn. (A.23). The position of the cone degenerate patch boundaries relative to the rod degenerate patch boundaries, and hence the question of whether the cone degenerate patch lies within or exceeds the rod degenerate patch, depends upon the value of f_{crit} , and is given by the following inequalities —

- when $f_{\text{crit}} \geq c_2/(2c_1)$, $\theta_{\text{crit}_1} \leq \theta_{r_1}$;
- when $f_{\text{crit}} < c_2/(2c_1)$, $\theta_{\text{crit}_1} > \theta_{r_1}$;
- when $f_{\text{crit}} \geq \bar{c}_2/(2\bar{c}_1)$, $\theta_{\text{crit}_2} \geq \theta_{r_2}$;
- when $f_{\text{crit}} < \bar{c}_2/(2\bar{c}_1)$, $\theta_{\text{crit}_2} < \theta_{r_2}$.

Thus, as would be expected intuitively, cone degenerate patches are wider for larger values of f_{crit} . For asymptotic analysis St-st Cases 2–8, see the Supplementary Materials.

Acknowledgements

P.A.R. is funded by BBSRC (BB/R014817/1) and thanks Tom Baden for allowing the time to pursue this research. P.A.R. also thanks the reviewers for their helpful and insightful comments.

References

- Aït-Ali, N., Fridlich, R., Millet-Puel, G., Clérin, E., Delalande, F., Jaillard, C., Blond, F., Perrocheau, L., Reichman, S., Byrne, L.C., Olivier-Bandini, A., Bellalou, J., Moyse, E., Bouillaud, F., Nicol, X., Dalkara, D., van Dorsselaer, A., Sahel, J.A., Lèveillard, T., 2015. Rod-derived cone viability factor promotes cone survival by stimulating aerobic glycolysis. *Cell* 161, 817–832. doi:<https://doi.org/10.1016/j.cell.2015.03.023>.
- Bender, C.M., Orszag, S.A., 1999. *Advanced Mathematical Methods for Scientists and Engineers I: Asymptotic Methods and Perturbation Theory*. Springer.
- Burns, J., Clarke, G., Lumsden, C.J., 2002. Photoreceptor death: Spatiotemporal patterns arising from one-hit death kinetics and a diffusible cell death factor. *Bull. Math. Biol.* 64, 1117–1145. doi:<https://doi.org/10.1006/bulm.2002.0320>.
- Byrne, L.C., Dalkara, D., Luna, G., Fisher, S.K., Clérin, E., Sahel, J.A., Lèveillard, T., Flannery, J.G., 2015. Viral-mediated RdCVF and RdCVFL expression protects cone and rod photoreceptors in retinal degeneration. *J. Clin. Invest.* 125, 105–116. doi:<https://doi.org/10.1172/JCI65654>.
- Camacho, E.T., Brager, D., Elachouri, G., Korneyeva, T., Millet-Puel, G., Sahel, J.A., Lèveillard, T., 2019. A mathematical analysis of aerobic glycolysis triggered by glucose uptake in cones. *Sci. Rep.* 9, 4162. doi:<https://doi.org/10.1038/s41598-019-39901-z>.
- Camacho, E.T., Colón Vélez, M.A., Hernández, D.J., Bernier, U.R., van Laarhoven, J., Wirkus, S., 2010. A mathematical model for photoreceptor interactions. *J. Theor. Biol.* 267, 638–646. doi:<https://doi.org/10.1016/j.jtbi.2010.09.006>.
- Camacho, E.T., Dobрева, A., Larripa, K., Rădulescu, A., Schmidt, D., Trejo, I., 2021. *Mathematical Modeling of Retinal Degeneration: Aerobic Glycolysis in a Single Cone*. Springer International Publishing, Cham. volume 22 of *Association for Women in Mathematics Series*. pp. 135–178. doi:https://doi.org/10.1007/978-3-030-57129-0_7.
- Camacho, E.T., Lenhart, S., Melara, L.A., Villalobos, M.C., Wirkus, S., 2020. Optimal control with MANF treatment of photoreceptor degeneration. *Math. Med. Biol.* 37, 1–21. doi:<https://doi.org/10.1093/imammb/dqz003>.

- Camacho, E.T., Léveillard, T., Sahel, J.A., Wirkus, S., 2016a. Mathematical model of the role of RdCVF in the coexistence of rods and cones in a healthy eye. *Bull. Math. Biol.* 78, 1394–1409. doi:<https://doi.org/10.1007/s11538-016-0185-x>.
- Camacho, E.T., Melara, L.A., Villalobos, M.C., Wirkus, S., 2014. Optimal control in the treatment of retinitis pigmentosa. *Bull. Math. Biol.* 76, 292–313. doi:<https://doi.org/10.1007/s11538-013-9919-1>.
- Camacho, E.T., Punzo, C., Wirkus, S.A., 2016b. Quantifying the metabolic contribution to photoreceptor death in retinitis pigmentosa via a mathematical model. *J. Theor. Biol.* 408, 75–87. doi:<https://doi.org/10.1016/j.jtbi.2016.08.001>.
- Camacho, E.T., Radulescu, A., Wirkus, S., 2016c. Bifurcation analysis of a photoreceptor interaction model for retinitis pigmentosa. *Commun. Nonlinear Sci. Numer. Simulat.* 38, 267–276. doi:<https://doi.org/10.1016/j.cnsns.2016.02.030>.
- Camacho, E.T., Wirkus, S., 2013. Tracing the progression of retinitis pigmentosa via photoreceptor interactions. *J. Theor. Biol.* 317, 105–118. doi:<https://doi.org/10.1016/j.jtbi.2012.09.034>.
- Chandler, M.J., Smith, P.J., Samuelson, D.A., Mackay, E.O., 1999. Photoreceptor density of the domestic pig retina. *Vet. Ophthalmol.* 2, 179–184. doi:<https://doi.org/10.1046/j.1463-5224.1999.00077.x>.
- Chrysostomou, V., Stone, J., Stowe, S., Barnett, N.L., Valter, K., 2008. The status of cones in the rhodopsin mutant P23H-3 retina: Light-regulated damage and repair in parallel with rods. *Invest. Ophthalmol. Vis. Sci.* 49, 1116–1125. doi:<https://doi.org/10.1167/iovs.07-1158>.
- Cideciyan, A.V., Hood, D.C., Huang, Y., Banin, E., Li, Z.Y., Stone, E.M., Milam, A.H., Jacobson, S.G., 1998. Disease sequence from mutant rhodopsin allele to rod and cone photoreceptor degeneration in man. *Proc. Natl. Acad. Sci.* 95, 7103–7108. doi:<https://doi.org/10.1073/pnas.95.12.7103>.
- Clarke, G., Collins, R.A., Leavitt, B.R., Andrews, D.F., Hayden, M.R., Lumsden, C.J., McInnes, R.R., 2000. A one-hit model of cell death in inherited neuronal degenerations. *Nature* 406, 195–199. doi:<https://doi.org/10.1038/35018098>.
- Clarke, G., Lumsden, C.J., McInnes, R.R., 2001. Inherited neurodegenerative diseases: the one-hit model of neurodegeneration. *Hum. Mol. Genet.* 10, 2269–2275. doi:<https://doi.org/10.1093/hmg/10.20.2269>.
- Clérin, E., Marussig, M., Sahel, J.A., Léveillard, T., 2020. Metabolic and redox signaling of the nucleoredoxin-like-1 gene for the treatment of genetic retinal diseases. *Int. J. Mol. Sci.* 21. doi:<https://doi.org/10.1155/2017/8475125>.
- Colón Vélez, M.A., Hernández, D.J., Bernier, U.R., van Laarhoven, J., Camacho, E.T., 2003. Mathematical Models for Photoreceptor Interactions. Technical Report. Cornell University, Department of Biological Statistics and Computational Biology.
- Curcio, C.A., Millican, C.L., Allen, K.A., Kalina, R.E., 1993. Aging of the human photoreceptor mosaic: evidence for selective vulnerability of rods in central retina. *Invest. Ophthalmol. Vis. Sci.* 34, 3278–3296.
- Curcio, C.A., Sloan, K.R., Kalina, R.E., Hendrickson, A.E., 1990. Human photoreceptor topography. *J. Comp. Neurol.* 292, 497–523. doi:<https://doi.org/10.1002/cne.902920402>.

- Daiger, S.P., Bowne, S.J., Sullivan, L.S., 2007. Perspective on genes and mutations causing retinitis pigmentosa. *Arch. Ophthalmol.* 125, 151–158. doi:10.1001/archophth.125.2.151.
- Dörrbaum, A.R., Kochen, L., Langer, J.D., Schuman, E.M., 2018. Local and global influences on protein turnover in neurons and glia. *eLife* 7, e34202. doi:10.7554/eLife.34202.
- Eden, E., Geva-Zatorsky, N., Issaeva, I., Cohen, A., Dekel, E., Danon, T., Cohen, L., Mayo, A., Alon, U., 2011. Proteome half-life dynamics in living human cells. *Science* 331, 764–768. doi:10.1126/science.1199784.
- Fintz, A.C., Audo, I., Hicks, D., Mohand-Saïd, S., Léveillard, T., Sahel, J., 2003. Partial characterization of retina-derived cone neuroprotection in two culture models of photoreceptor degeneration. *Invest. Ophthalmol. Vis. Sci.* 44, 818–825. doi:https://doi.org/10.1167/iovs.01-1144.
- Gaillard, F., Kuny, S., Sauv e, Y., 2009. Topographic arrangement of S-cone photoreceptors in the retina of the diurnal Nile grass rat (*Arvicanthis niloticus*). *Invest. Ophthalmol. Vis. Sci.* 50, 5426–5434. doi:https://doi.org/10.1167/iovs.09-3896.
- García-Ayuso, D., Ortín-Martínez, A., Jiménez-López, M., Galindo-Romero, C., Cuenca, N., Pinilla, I., Vidal-Sanz, M., Agudo-Barriuso, M., Villegas-Pérez, M.P., 2013. Changes in the photoreceptor mosaic of P23H-1 rats during retinal degeneration: Implications for rod-cone dependent survival. *Invest. Ophthalmol. Vis. Sci.* 54, 5888–5900. doi:https://doi.org/10.1167/iovs.13-12643.
- Grover, S., Fishman, G.A., Brown Jr, J., 1998. Patterns of visual field progression in patients with retinitis pigmentosa. *Ophthalmology* 105, 1069–1075. doi:https://doi.org/10.1016/S0161-6420(98)96009-2.
- Guérin, C.J., Lewis, G.P., Fisher, S.K., Anderson, D.H., 1993. Recovery of photoreceptor outer segment length and analysis of membrane assembly rates in regenerating primate photoreceptor outer segments. *Invest. Ophthalmol. Vis. Sci.* 34, 175–183.
- Gupta, N., Brown, K.E., Milam, A.H., 2003. Activated microglia in human retinitis pigmentosa, late-onset retinal degeneration, and age-related macular degeneration. *Exp. Eye Res.* 76, 463–471. doi:https://doi.org/10.1016/S0014-4835(02)00332-9.
- Hamel, C., 2006. Retinitis pigmentosa. *Orphanet. J. Rare Dis.* 1, 40. doi:https://doi.org/10.1186/1750-1172-1-40.
- Hartong, D.T., Berson, E.L., Dryja, T.P., 2006. Retinitis pigmentosa. *Lancet* 368, 1795–1809. doi:https://doi.org/10.1016/S0140-6736(06)69740-7.
- Ji, Y., Zhu, C.L., Grzywacz, N.M., Lee, E.J., 2012. Rearrangement of the cone mosaic in the retina of the rat model of retinitis pigmentosa. *J. Comp. Neurol.* 520, 874–888. doi:https://doi.org/10.1002/cne.22800.
- Jonnal, R.S., Besecker, J.R., Derby, J.C., Kocaoglu, O.P., Cense, B., Gao, W., Wang, Q., Miller, D.T., 2010. Imaging outer segment renewal in living human cone photoreceptors. *Opt. Express* 18, 5257–5270. doi:https://doi.org/10.1364/OE.18.005257.
- Jonnal, R.S., Kocaoglu, O.P., Wang, Q., Lee, S., Miller, D.T., 2012. Phase-sensitive imaging of the outer retina using optical coherence tomography and adaptive optics. *Biomed. Opt. Express* 3, 104–124. doi:https://doi.org/10.1364/BOE.3.000104.

- Jürgens, K.D., Peters, T., Gros, G., 1994. Diffusivity of myoglobin in intact skeletal muscle cells. *Proc. Natl. Acad. Sci.* 91, 3829–3833. doi:<https://doi.org/10.1073/pnas.91.9.3829>.
- Kocaoglu, O.P., Liu, Z., Zhang, F., Kurokawa, K., Jonnal, R.S., Miller, D.T., 2016. Photoreceptor disc shedding in the living human eye. *Biomed. Opt. Express* 7, 4554–4568. doi:<https://doi.org/10.1364/BOE.7.004554>.
- Lee, D.C., Vazquez-Chona, F.R., Ferrell, W.D., Tam, B.M., Jones, B.W., Marc, R.E., Moritz, O.L., 2012. Dysmorphic photoreceptors in a P23H mutant rhodopsin model of retinitis pigmentosa are metabolically active and capable of regenerating to reverse retinal degeneration. *J. Neurosci.* 32, 2121–2128. doi:<https://doi.org/10.1523/JNEUROSCI.4752-11.2012>.
- Lee, E.J., Ji, Y., Zhu, C.L., Grzywacz, N.M., 2011. Role of Müller cells in cone mosaic rearrangement in a rat model of retinitis pigmentosa. *Glia* 59, 1107–1117. doi:<https://doi.org/10.1002/glia.21183>.
- Léveillard, T., Aït-Ali, N., 2017. Cell signaling with extracellular thioredoxin and thioredoxin-like proteins: Insight into their mechanisms of action. *Oxid. Med. Cell. Longev.* 2017. doi:<https://doi.org/10.1155/2017/8475125>.
- Léveillard, T., Mohand-Saïd, S., Lorentz, O., Hicks, D., Fintz, A.C., Clérin, E., Simonutti, M., Forster, V., Cavusoglu, N., Chalmel, F., Dollé, P., Poch, O., Lambrou, G., Sahel, J.A., 2004. Identification and characterization of rod-derived cone viability factor. *Nat. Genet.* 36, 755–759. doi:<https://doi.org/10.1038/ng1386>.
- Léveillard, T., Sahel, J.A., 2010. Rod-derived cone viability factor for treating blinding diseases: From clinic to redox signaling. *Sci. Transl. Med.* 2, ps16. doi:<https://doi.org/10.1126/scitranslmed.3000866>.
- Léveillard, T., Sahel, J.A., 2017. Metabolic and redox signaling in the retina. *Cell. Mol. Life Sci.* 74, 3649–3665. doi:<https://doi.org/10.1007/s00018-016-2318-7>.
- Mei, X., Chaffiol, A., Kole, C., Yang, Y., Millet-Puel, G., Clérin, E., Aït-Ali, N., Bennett, J., Dalkara, D., Sahel, J.A., Duebel, J., Léveillard, T., 2016. The thioredoxin encoded by the rod-derived cone viability factor gene protects cone photoreceptors against oxidative stress. *Antioxidants & Redox Signaling* 24, 909–923. doi:<https://doi.org/10.1089/ars.2015.6509>.
- Mervin, K., Stone, J., 2002. Developmental death of photoreceptors in the C57BL/6J mouse: Association with retinal function and self-protection. *Exp. Eye Res.* 75, 703–713. doi:<https://doi.org/10.1006/exer.2002.2063>.
- Mohand-Saïd, S., Deudon-Combe, A., Hicks, D., Simonutti, M., Forster, V., Fintz, A.C., Léveillard, T., Dreyfus, H., Sahel, J.A., 1998. Normal retina releases a diffusible factor stimulating cone survival in the retinal degeneration mouse. *Proc. Natl. Acad. Sci.* 95, 8357–8362. doi:<https://doi.org/10.1073/pnas.95.14.8357>.
- Mohand-Saïd, S., Hicks, D., Dreyfus, H., Sahel, J.A., 2000. Selective transplantation of rods delays cone loss in a retinitis pigmentosa model. *Arch. Ophthalmol.* 118, 807–811. doi:[10.1001/archophth.118.6.807](https://doi.org/10.1001/archophth.118.6.807).
- Mohand-Saïd, S., Hicks, D., Léveillard, T., Picaud, S., Porto, F., Sahel, J.A., 2001. Rod-cone interactions: Developmental and clinical significance. *Prog. Retin. Eye Res.* 20, 451–467. doi:[https://doi.org/10.1016/S1350-9462\(01\)00006-4](https://doi.org/10.1016/S1350-9462(01)00006-4).

- Mohand-Saïd, S., Hicks, D., Simonutti, M., Tran-Minh, D., Deudon-Combe, A., Dreyfus, H., Silverman, M.S., Ogilvie, J.M., Tenkova, T., Sahel, J., 1997. Photoreceptor transplants increase host cone survival in the retinal degeneration (rd) mouse. *Ophthalmic Res.* 29, 290–297. doi:<https://doi.org/10.1159/000268027>.
- Musarella, M.A., MacDonald, I.M., 2011. Current concepts in the treatment of retinitis pigmentosa. *J. Ophthalmol.* 2011, 753547. doi:<https://doi.org/10.1155/2011/753547>.
- Ortín-Martínez, A., Nadal-Nicolás, F.M., Jiménez-López, M., Alburquerque-Béjar, J.J., Nieto-López, L., García-Ayuso, D., Villegas-Pérez, M.P., Vidal-Sanz, M., Agudo-Barriuso, M., 2014. Number and distribution of mouse retinal cone photoreceptors: Differences between an albino (Swiss) and a pigmented (C57/BL6) strain. *PLoS One* 9, e102392. doi:<https://doi.org/10.1371/journal.pone.0102392>.
- Oyster, C.W., 1999. *The Human Eye: Structure and Function*. Sinauer Associates Inc.
- Pircher, M., Kroisamer, J.S., Felberer, F., Sattmann, H., Götzinger, E., Hitznerberger, C.K., 2011. Temporal changes of human cone photoreceptors observed in vivo with SLO/OCT. *Biomed. Opt. Express* 2, 100–112. doi:<https://doi.org/10.1364/BOE.2.000100>.
- Punzo, C., Kornacker, K., Cepko, C.L., 2009. Stimulation of the insulin/mTOR pathway delays cone death in a mouse model of retinitis pigmentosa. *Nat. Neurosci.* 12, 44–52. doi:<https://doi.org/10.1038/nn.2234>.
- Punzo, C., Xiong, W., Cepko, C.L., 2012. Loss of daylight vision in retinal degeneration: Are oxidative stress and metabolic dysregulation to blame? *J. Biol. Chem.* 287, 1642–1648. doi:<https://doi.org/10.1074/jbc.R111.304428>.
- Reichman, S., Kalathur, R.K.R., Lambard, S., Ait-Ali, N., Yang, Y., Lardenois, A., Ripp, R., Poch, O., Zack, D.J., Sahel, J.A., Léveillard, T., 2010. The homeobox gene CHX10/VSX2 regulates RdCVF promoter activity in the inner retina. *Hum. Mol. Genet.* 19, 250–261. doi:<https://doi.org/10.1093/hmg/ddp484>.
- Ripps, H., 2002. Cell death in retinitis pigmentosa: Gap junctions and the ‘bystander’ effect. *Exp. Eye Res.* 74, 327–336. doi:<https://doi.org/10.1006/exer.2002.1155>.
- Roberts, P.A., Gaffney, E.A., Luthert, P.J., Foss, A.J.E., Byrne, H.M., 2016a. Mathematical and computational models of the retina in health, development and disease. *Prog. Retin. Eye Res.* 53, 48–69. doi:<https://doi.org/10.1016/j.preteyeres.2016.04.001>.
- Roberts, P.A., Gaffney, E.A., Luthert, P.J., Foss, A.J.E., Byrne, H.M., 2016b. Retinal oxygen distribution and the role of neuroglobin. *J. Math. Biol.* 73, 1–38. doi:<https://doi.org/10.1007/s00285-015-0931-y>.
- Roberts, P.A., Gaffney, E.A., Luthert, P.J., Foss, A.J.E., Byrne, H.M., 2017. Mathematical models of retinitis pigmentosa: The oxygen toxicity hypothesis. *J. Theor. Biol.* 425, 53–71. doi:<https://doi.org/10.1016/j.jtbi.2017.05.006>.
- Roberts, P.A., Gaffney, E.A., Whiteley, J.P., Luthert, P.J., Foss, A.J.E., Byrne, H.M., 2018. Predictive mathematical models for the spread and treatment of hyperoxia-induced photoreceptor degeneration in retinitis pigmentosa. *Invest. Ophthalmol. Vis. Sci.* 59, 1238–1249. doi:<https://doi.org/10.1167/iovs.17-23177>.

- Shintani, K., Shechtman, D.L., Gurwood, A.S., 2009. Review and update: Current treatment trends for patients with retinitis pigmentosa. *Optometry* 80, 384–401. doi:<https://doi.org/10.1016/j.optm.2008.01.026>.
- Sieving, P.A., Caruso, R.C., Tao, W., Coleman, H.R., Thompson, D.J.S., Fullmer, K.R., Bush, R.A., 2006. Ciliary neurotrophic factor (CNTF) for human retinal degeneration: Phase I trial of CNTF delivered by encapsulated cell intraocular implants. *Proc. Natl. Acad. Sci.* 103, 3896–3901. doi:<https://doi.org/10.1073/pnas.0600236103>.
- Stone, J., Maslim, J., Valter-Kocsi, K., Mervin, K., Bowers, F., Chu, Y., Barnett, N., Provis, J., Lewis, G., Fisher, S.K., Bistid, S., Gargini, C., Cervetto, L., Merin, S., Pe'er, J., 1999. Mechanisms of photoreceptor death and survival in mammalian retina. *Prog. Retin. Eye Res.* 18(6), 689–735. doi:[https://doi.org/10.1016/S1350-9462\(98\)00032-9](https://doi.org/10.1016/S1350-9462(98)00032-9).
- Tao, W., Wen, R., Goddard, M.B., Sherman, S.D., O'Rourke, P.J., Stabila, P.F., Bell, W.J., Dean, B.J., Kauper, K.A., Budz, V.A., Tsiaras, W.G., Acland, G.M., Pearce-Kelling, S., Laties, A.M., Aguirre, G.D., 2002. Encapsulated cell-based delivery of CNTF reduces photoreceptor degeneration in animal models of retinitis pigmentosa. *Invest. Ophthalmol. Vis. Sci.* 43, 3292–3298.
- Travis, G.H., Sutcliffe, J.G., Bok, D., 1991. The retinal degeneration slow (rds) gene product is a photoreceptor disc membrane-associated glycoprotein. *Neuron* 6, 61–70. doi:[https://doi.org/10.1016/0896-6273\(91\)90122-G](https://doi.org/10.1016/0896-6273(91)90122-G).
- Valter, K., Maslim, J., Bowers, F., Stone, J., 1998. Photoreceptor dystrophy in the RCS rat: roles of oxygen, debris, and bFGF. *Invest. Ophthalmol. Vis. Sci.* 39, 2427–2442.
- Wifvat, K., Camacho, E.T., Wirkus, S., Léveillard, T., 2021. The role of RdCVFL in a mathematical model of photoreceptor interactions. *J. Theor. Biol.* 520, 110642. doi:<https://doi.org/10.1016/j.jtbi.2021.110642>.
- Yang, Y., Mohand-Said, S., Danan, A., Simonutti, M., Fontaine, V., Clérin, E., Picaud, S., Leveillard, T., Sahel, J.A., 2009. Functional cone rescue by RdCVF protein in a dominant model of retinitis pigmentosa. *Mol. Ther.* 17, 787–795. doi:<https://doi.org/10.1038/mt.2009.28>.
- Young, R.W., 1967. The renewal of photoreceptor cell outer segments. *J. Cell Biol.* 33, 61–72. doi:<https://doi.org/10.1083/jcb.33.1.61>.
- Young, R.W., 1971. The renewal of rod and cone outer segments in the rhesus monkey. *J. Cell Biol.* 49, 303–318. doi:<https://doi.org/10.1083/jcb.49.2.303>.
- Young, R.W., 1978. The daily rhythm of shedding and degradation of rod and cone outer segment membranes in the chick retina. *Invest. Ophthalmol. Vis. Sci.* 17, 105–116.
- Young, R.W., Bok, D., 1969. Participation of the retinal pigment epithelium in the rod outer segment renewal process. *J. Cell Biol.* 42, 392–403. doi:<https://doi.org/10.1083/jcb.42.2.392>.
- Zhang, C.J., Ma, Y., Jin, Z.B., 2021. The road to restore vision with photoreceptor regeneration. *Exp. Eye Res.* 202, 108283. doi:<https://doi.org/10.1016/j.exer.2020.108283>.
- Zhu, C.L., Ji, Y., Lee, E.J., Grzywacz, N.M., 2013. Spatiotemporal pattern of rod degeneration in the S334ter-line-3 rat model of retinitis pigmentosa. *Cell Tissue Res.* 351, 29–40. doi:<https://doi.org/10.1007/s00441-012-1522-5>.